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3.1.4. –

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1.1	..	9
1.2	15
1.3	...	20
2	28
3	37
4	42
4.1	42
4.2	48
4.3	50
5	62
5.1	62
5.2	-	
	65
5.3		
,	69
6	86
	93
	94
	95
	97

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 ,
 (. ., 2014; . . ., 2019; . . ., 2019; . . ., 2020).
 7–
 11%, 70–87% (. ., 2014; Park H.J. et al., 2016).
 68% (. ., . ., 2016; Kannar V., Lingaiah H.K., Sunita V., 2012), – 60%,
 – 64,2–86,7% (. ., . ., 2013).
 (. ., . ., 2012; . ., . ., 2017; . . ., 2019).
 (. ., . ., 2015; . . ., 2019).
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 , (. ., 2013; . ., . ., 2015; Cicinelli E. et al., 2015).
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 (. ., 2013; Cicinelli E. et al., 2015; Moreno I. et al., 2018).

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VEGF

TGF-

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VEGF

TGF-

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– : Status Praesens, 2021. – 68 .

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on reproductive medicine XV international ongress» (,

2021); XIII - :

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[121]. -

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8 72% [163, 170]. , -

57% [135].

14–67,5% [102, 121, 122].

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13% [137]. , -

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[102]. , -

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[49]. -

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[135, 177].

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[138, 181].

(VEGF),

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[134].

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[188]. « » , -
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 VEGF, -
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 [136]. VEGF ,
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 VEGF , -
 (RIF) [171]. -
 F.M. Carvalho
 . [141].
 X. Chen . [160], 85,7% -
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 7,3% . , -
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 [160]. - , -

[166].

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[60, 181].

VEGF

[61].

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[75],

, VEGF.

[115].

S. Tandulwadkar

. [113]

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VEGF

[87].

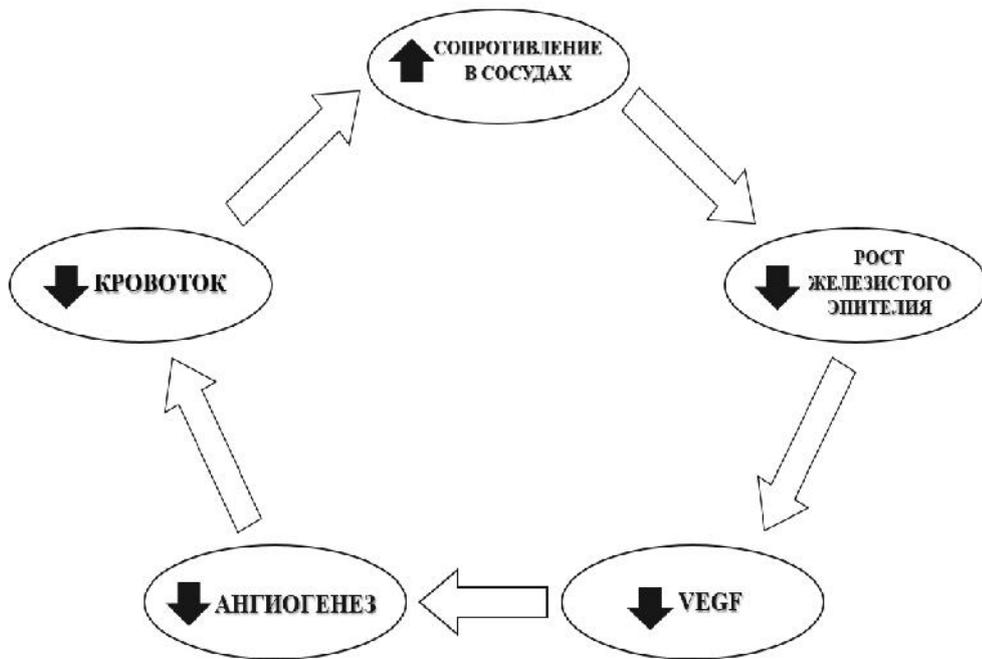
VEGF [146].

VEGF-A,

[95].

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VEGF,
[37, 138]. VEGF

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VEGF

VEGF

TGF- 1(1)

TGF-b1 (0,483 0,214
 0,260 0,208), MMP-9, -
 (0,190 0,106 0,234 0,096). -
 . TGF-b1 -
 TGF-b1 / Smad
 . TGF-b1 ,
 MMP-9 [112,
 180]. TGF 1 -
 (VSMC), -
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 , TGF 1 ,
 [167]. TGF- -
 [151, 167]. TGF- -
 [155] [75].
 S. Rajaei . [125] , -
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 - [172],
 (-
) [127].

1.2

[5, 11, 62, 127].

[82].

[154].

[122].

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[85].

(2007) [80].

(, ,).

(H&E)

CD138,

[149, 153, 182].

Syndecan-1-CD138

25–30% [43].

CD138

48,3%

8000 [20]. . . . [96] -

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-), CD 20+ (-), CD 138+ (), CD 4+ -

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×400, 4 ().

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[96]. -

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7-10 [83, 92]. , -

19-22 . . . 28- , -

[96]. -

p16INK4a [16, 17, 56, 57, 94]. -

[35, 47, 74, 185]. -

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() [104]. -

(2190) , -

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[147].

(D 138) [40, 63, 90]. . . [7]

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(11 ,
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[65]. , -

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7 [14, 29];

2020 . 8 [110, 161]. V. Pinto . [107] -

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Computer-aided AnaLysis).
 (FI), (VFI), (VI)
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(2007)
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 [71, 77].

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[50, 91].

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[39, 70, 72].

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[30, 90, 118]

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-6, , -
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, CD95+, CD4+ CD8+ -
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, HLA-DRII, -
TLR4, -a, HBD1 [48]. -
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[13, 34,
66, 101].

(80–160 200 5%
) [2], , [2], , -
[53]. -
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[8, 18, 99]. . . [46]
(4–6 2–3
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 (20 2) [12, 32, 84].
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 [139, 144, 156, 173], -
 [105, 106, 117, 133, 183].
 E. Papanikolaou . [140],
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 150 7 [140]. -
 , [126, 129, 145, 150]. -
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 (VEGF) [179].
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 [132].
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(9,8 8 ; <0,0001).

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[45, 168].

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«Horny Goat Weed»

Yin Yang Huo .

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[130, 148, 176, 178].

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[143].

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[64].

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[175]. 2015 Y. Chang .

[114] « »

PRP (platelet-rich plasm)- - , -

PRP- . -

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150 000 400 000 . -

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[84].

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2. PDGF- -

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3. VEGF-

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[109, 111]. PRP

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[58, 124]. 2019

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[152].

PRP

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[59,

51, 81, 131].

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PRP

[108, 116, 128,

184, 187].

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PRP

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[174].

PRP-

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[159].

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8 PRP 70–77% (p<0,05);
– 97% (<0,01). [162] -

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 (28))
 (10 12) [55, 64].

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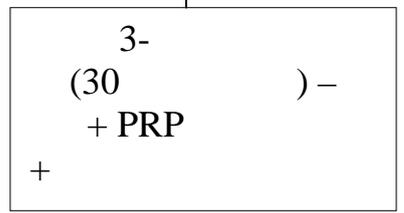
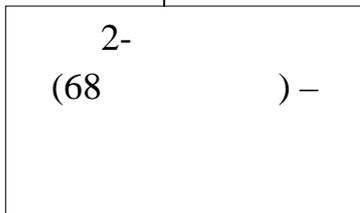
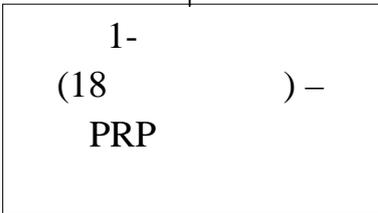
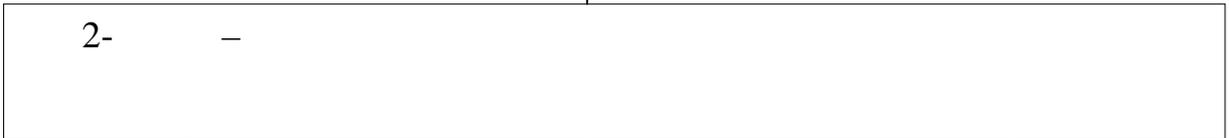
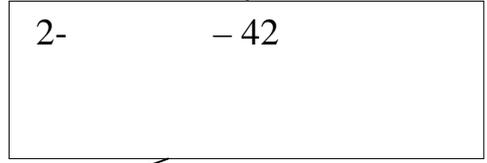
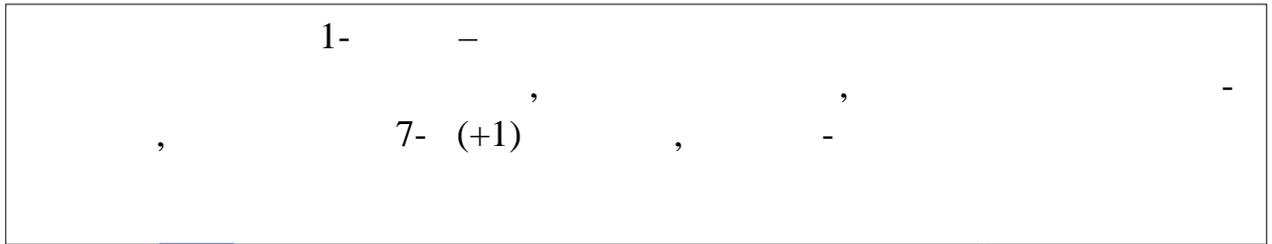
- « »

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 3- - 30 , -
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– «real-time processing»).

0,3–0,7

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(2007) (1).

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(M±m)

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	0,59±0,01
	0,48±0,02
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 (H-score), (1).

$$HS = 1a + 2b + 3c, \quad (1)$$

: - % , b - % ,
 - % , 1, 2, 3 - , -

- 0-10 - ;
 - 11-100 - ;

- 101-200 -

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- 201-300 -

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CD138 (,)

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CD138

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Hscore = Pi·i,

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VEGF, TGF-

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H-score,

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- 11-100 -

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- 101-200 -

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- 201-300 -

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MS Excel.

«Microsoft Excel» «Statistica 10.0».

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(10 (12 (3- : 17-b 0,1%- (28)) -1 5- 25-).

, , , . , - , [84]. , [84]. - , , , , . , [165], , . « » (,). . , , .

[6, 23, 33].

8 « »
(,).

4 / [66, 70].

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3 [22, 97],

– 1,5 / [25].

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[143].

103],

[68,

[175].

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(Pure-PRP(P-PRP)), L-PRP, (P-PRF)
(L-PRF).

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- «Cortexil» (3), -

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9 «Cortexil» (4) 620g,
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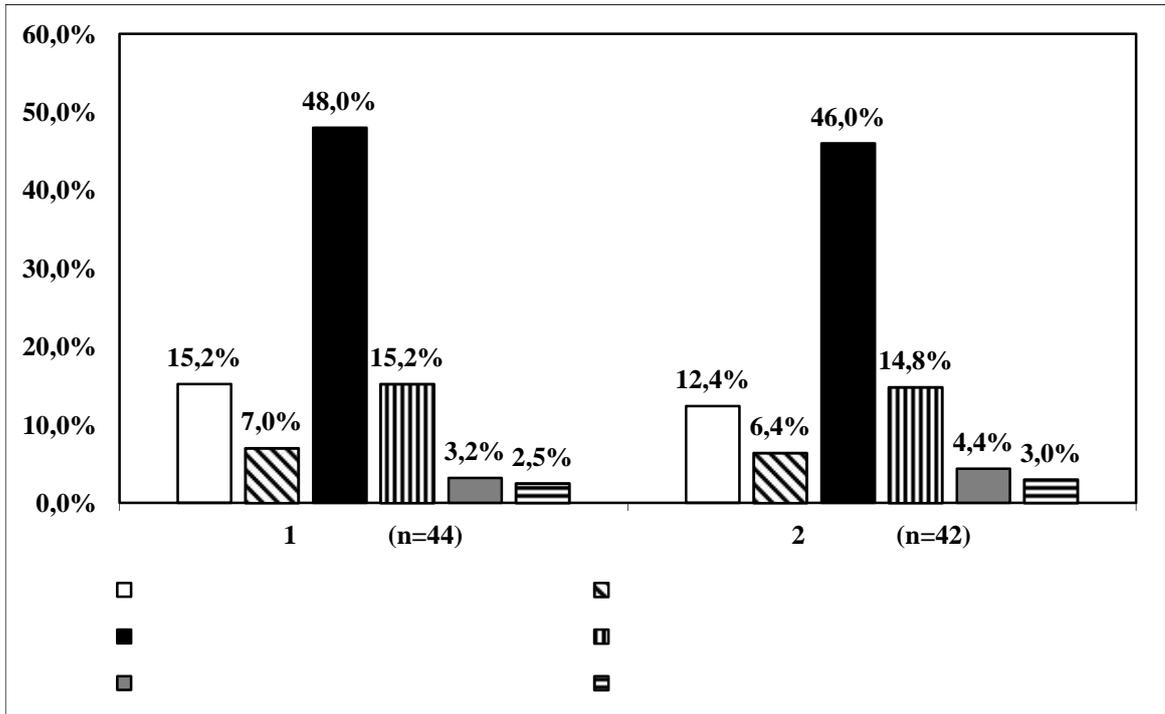
“Buffy Coat”

“Buffy Coat” –

PRP 10 9 1 3-4

PRP. PRP [4, 28].

5.



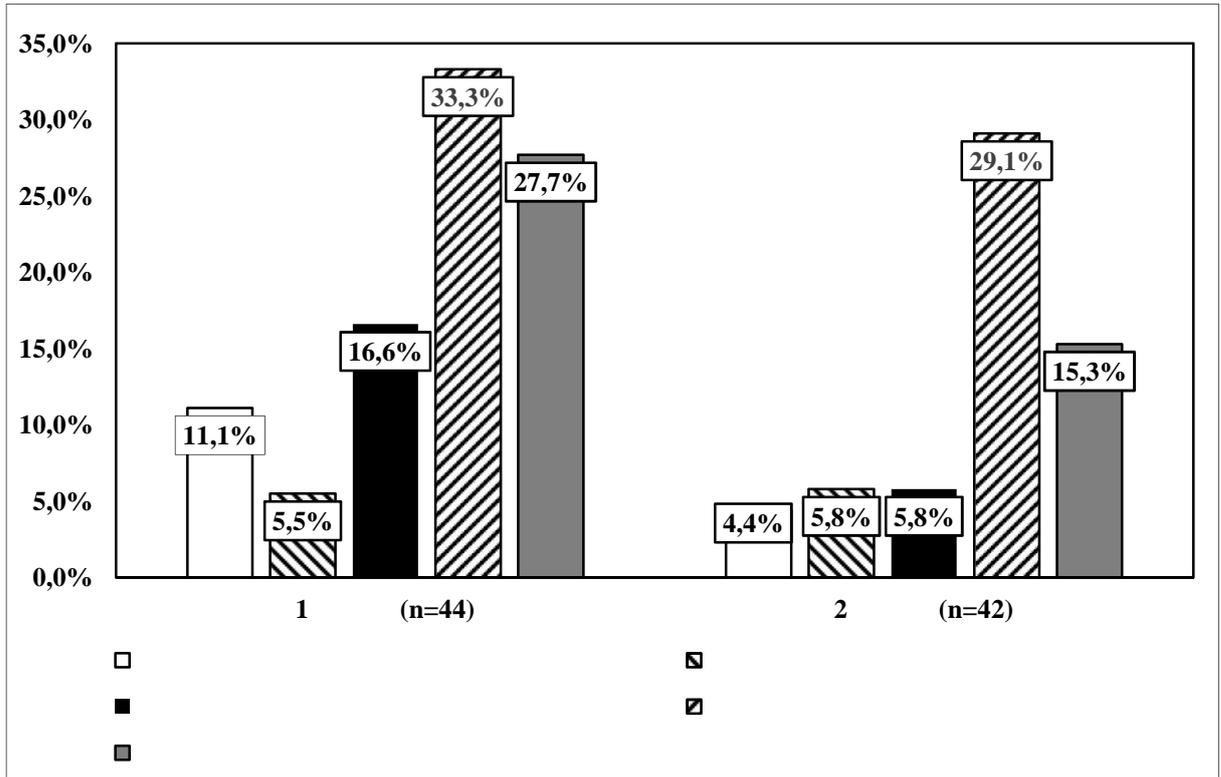
5 –

1- , 2- .

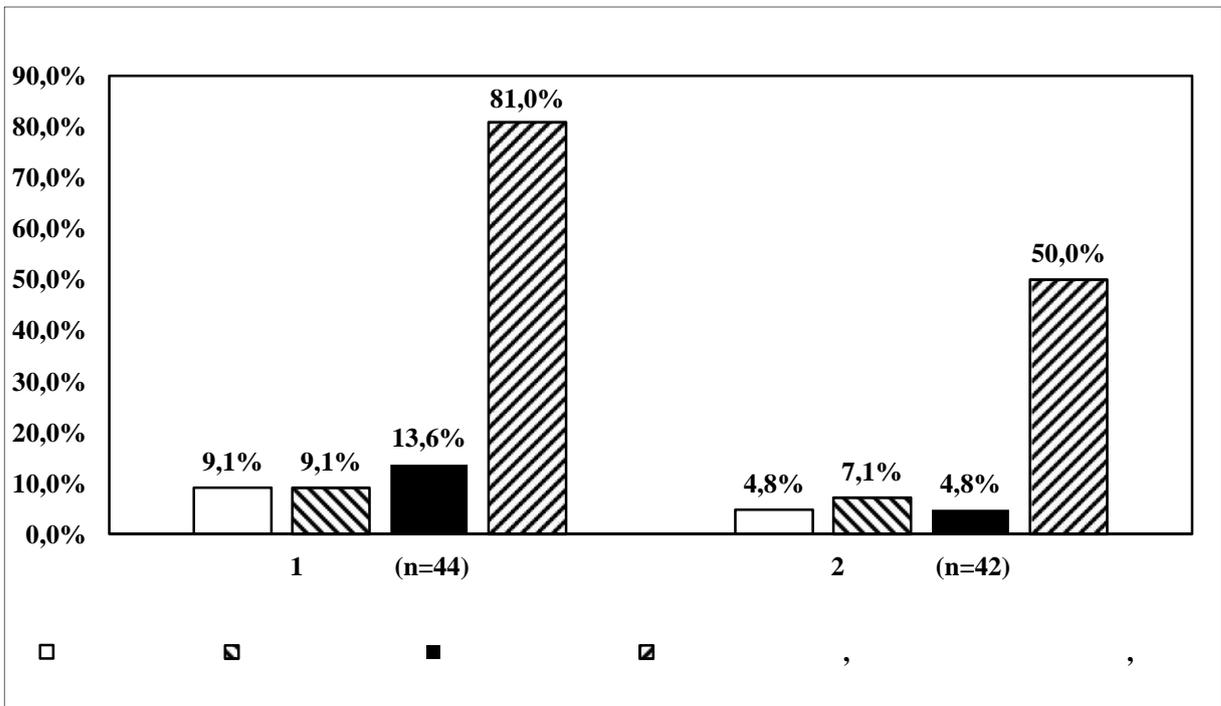
6.

() : – 80,95% ,

7.



6 -



7 -

1(0; 3),
 - 0 (0; 2).
 2 (1; 2), - 0 (0; 2).
 2 (1; 5),
 2 (1; 6), (>0,05).
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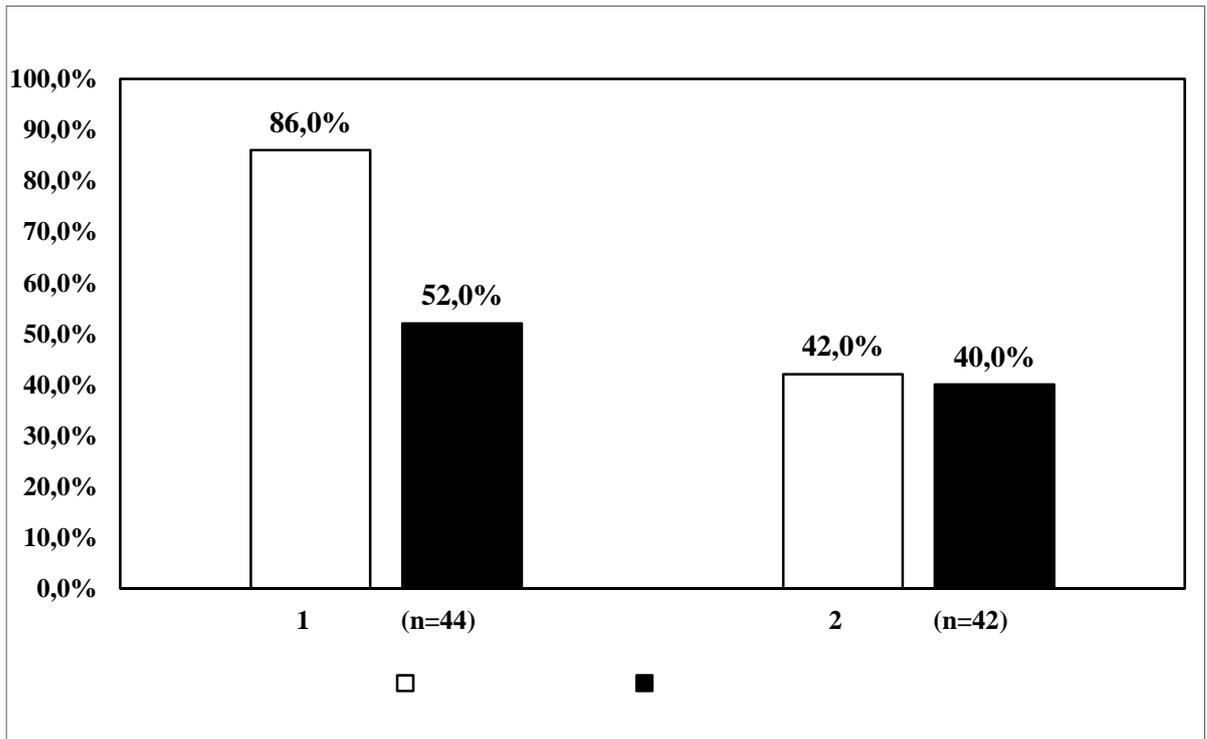
2-

	1- (n=44)	2- (n=42)
	2 [0; 5]	2 [1; 4]
	1 [0; 3]	0 [0; 2]
	2 [1; 2]	0 [0; 2]
	2 [1; 5]	2 [1; 6]

(>0,05).

13 [12; 15] , 13 [10; 15],
 28 [24; 34] , 28 [22; 32].

86%, 2- , 1-
 52% 1- 42% .
 40% 2-
 (8).



8 –

(>0,05).

3.

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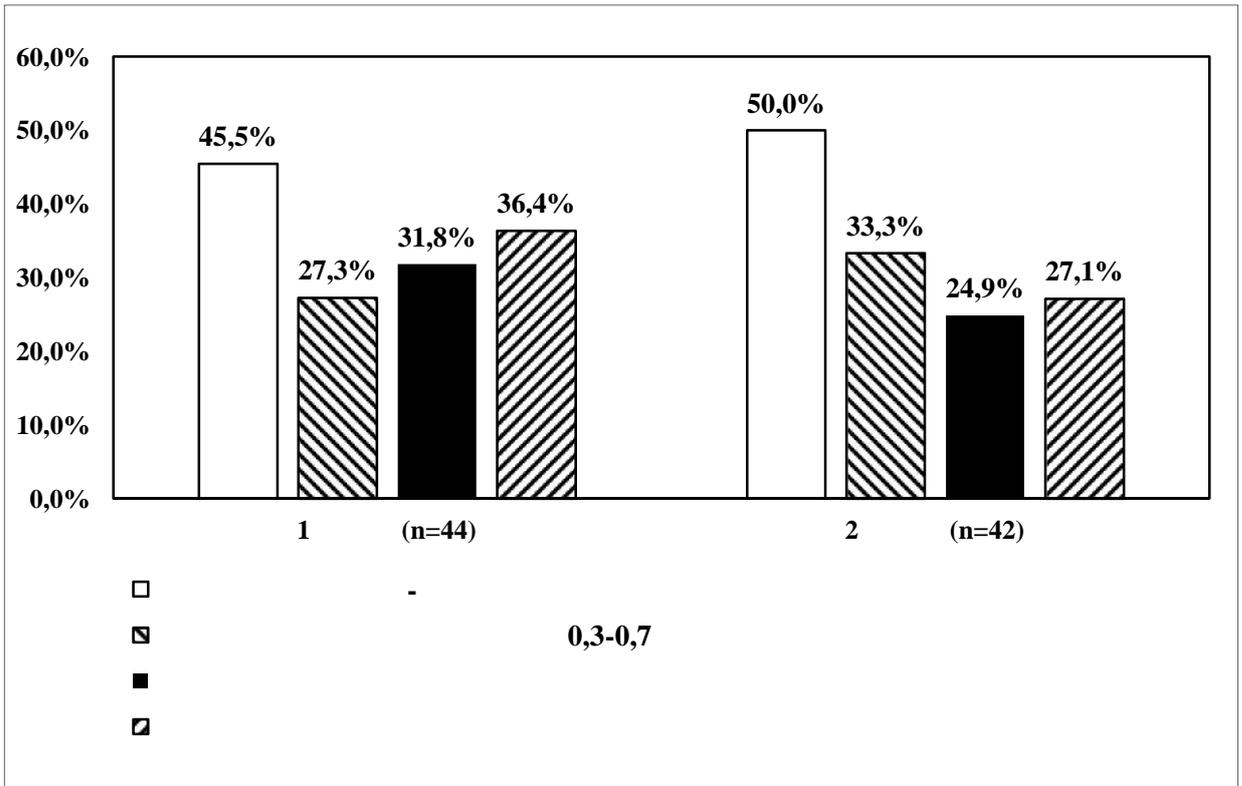
	1- (n=44)	2- (n=42)
1	2	3
, 10 ⁹ /	4,48 [2,4; 9,7]	4,05 [3; 8,02]

3		
1	2	3
, 10 ⁹ /	285 [212; 340]	248 [214; 345]
, 10 ¹² /	4,74 [4,45; 5,67]	4,86 [3,78; 5,82]
3		
, /	125,45 [112,74; 144,74]	128,85 [112; 141]
, /	75,4 [56; 86]	68,7 [64; 80]
, /	4,69 [3,9; 5,64]	4,37 [3,1; 5,4]
, /	15 [8; 24]	16 (4; 22)
, /	16 [10; 24]	16 [8; 20]
, /	3,4 [2,6; 4,5]	3,6 [2,5; 6,5]
, %	102 [100; 106]	101 [95; 110]
,	29,3 [24,8; 32,4]	29,2 [20,8; 40,4]
– , /	332 [229; 440]	330 [228; 454]

4.2

7- (+1)

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- 3- 4-

(<0,05).

49

5,4 (3,8; 7,7) . - 3,2 (2,3; 6,8) , -

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4 –

	1- (n=44)	2- (n=42)
,	3,2 [2,3; 6,8]	5,4 [3,8; 7,7]

« » -

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5 –

()	1- (n=44)	2- (n=42)
	0,88 [0,76; 0,94]	0,86 [0,72;0,96]
	0,80 [0,70; 0,90]	0,80 [0,70;0,90]
	0,67 [0,56; 0,80]	0,72 [0,62;0,82]
	0,68 [0,54; 0,75]	0,67 [0,54;0,76]
	0,20 [0; 0,64]	0,48 [0,20;0,67]
	0 [0; 0]	0,47 [0,20;0,60]

0,47 [0,20; 0,60]. -

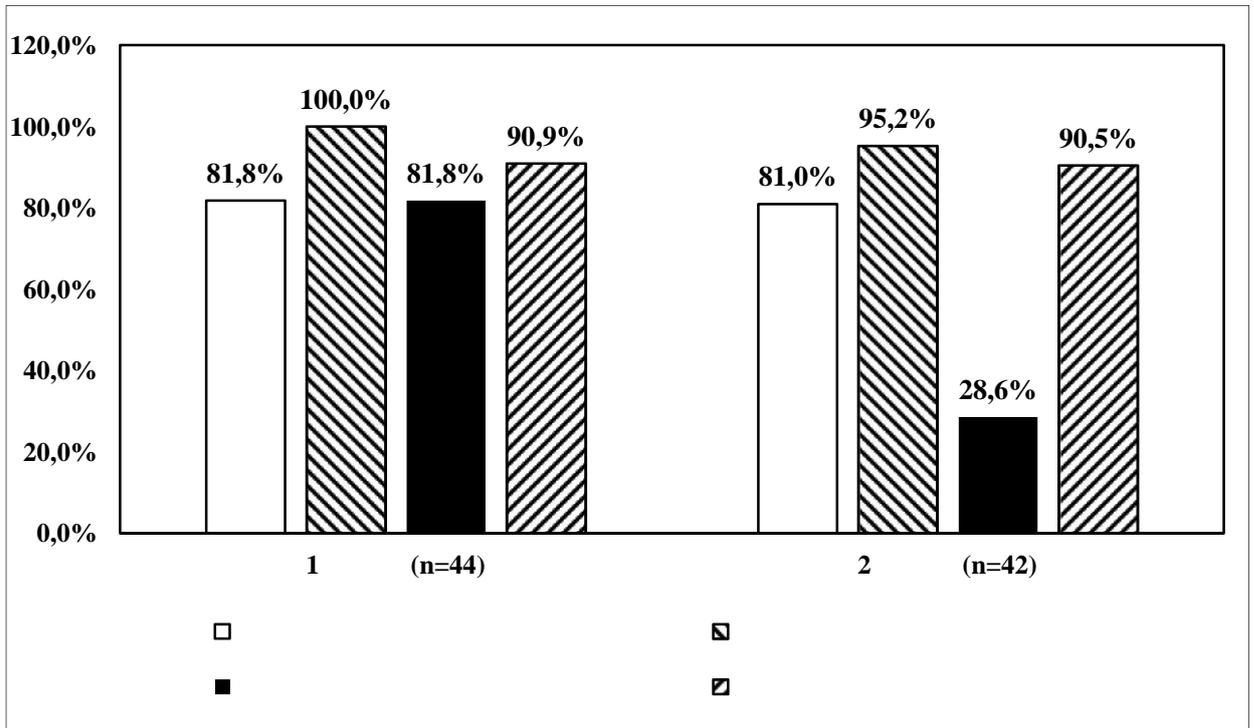
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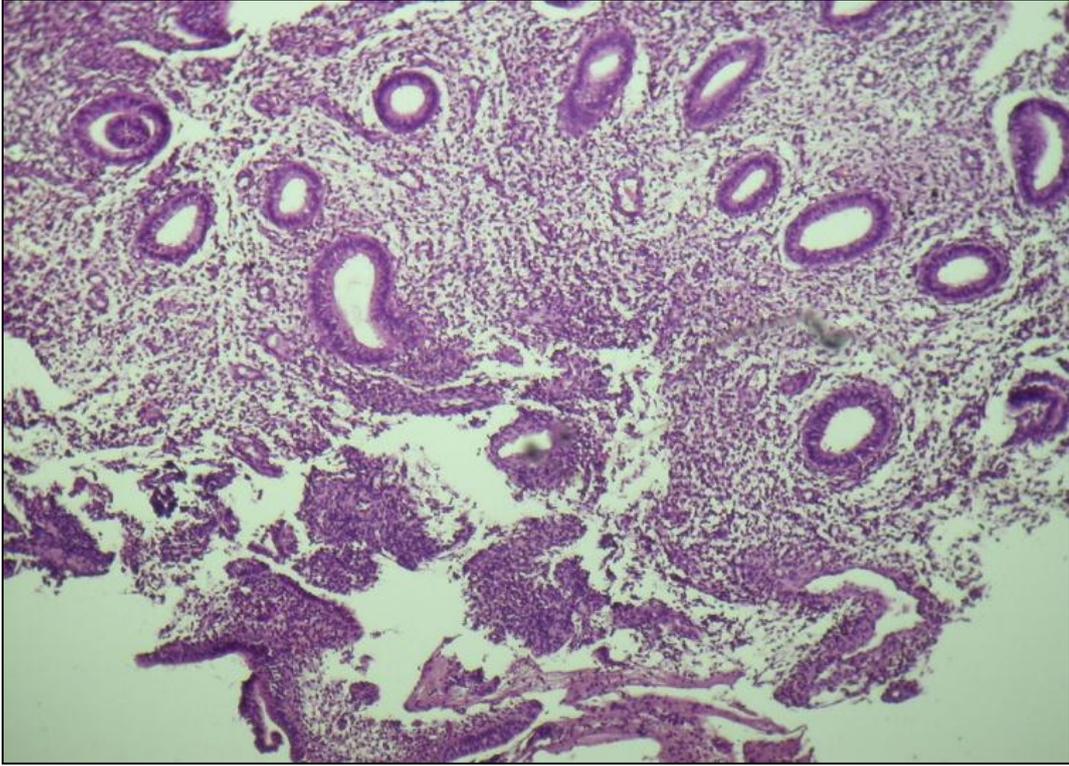
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95,2% , (11).



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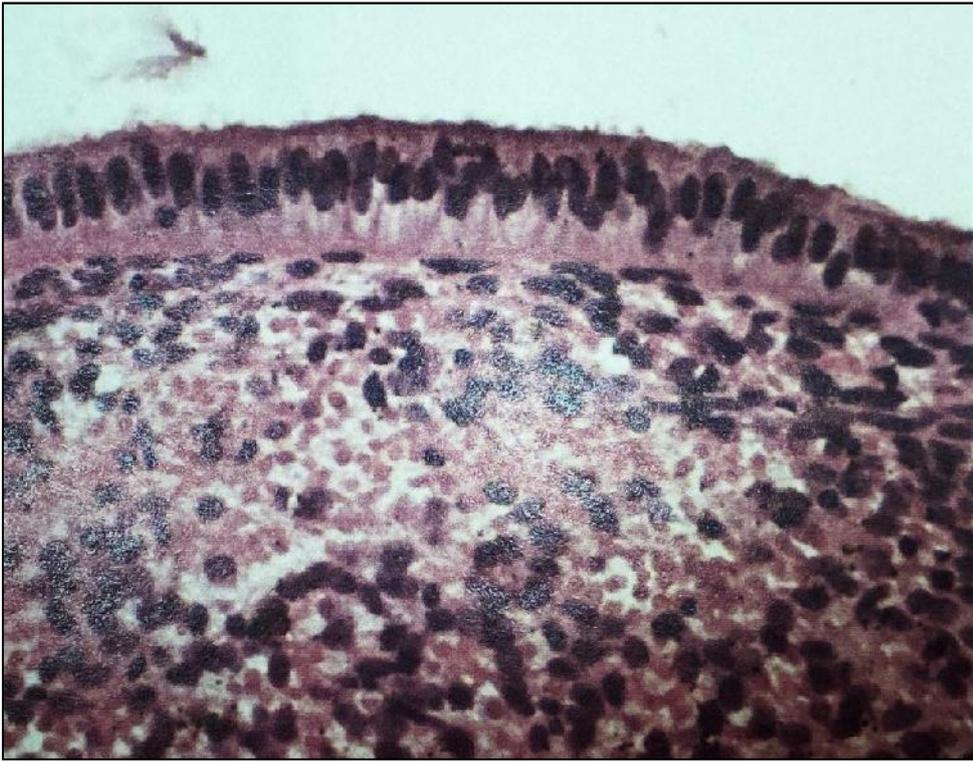


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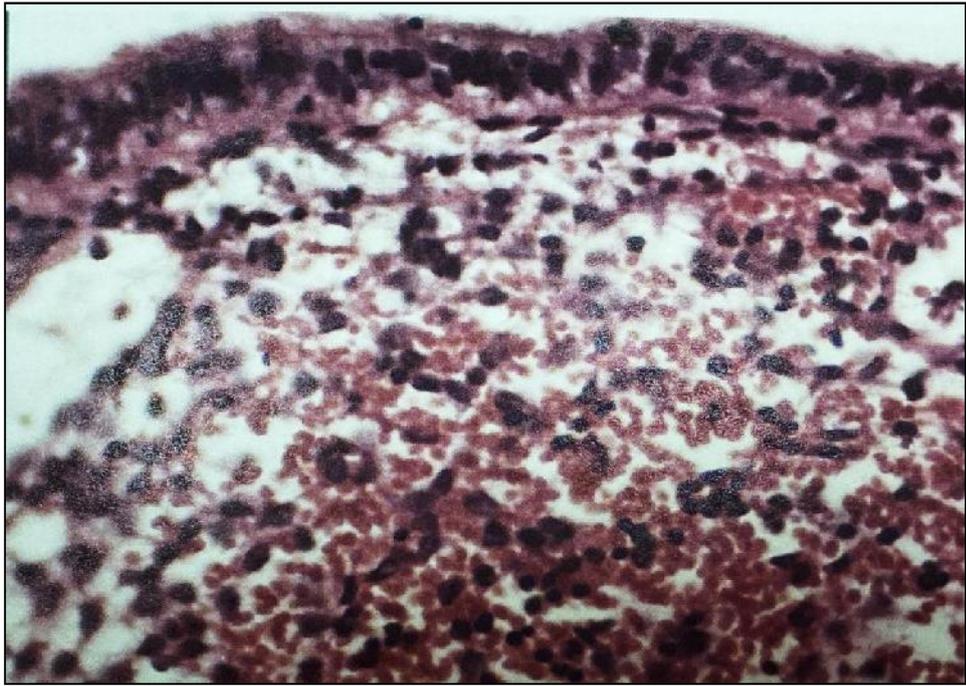
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(15, 16).

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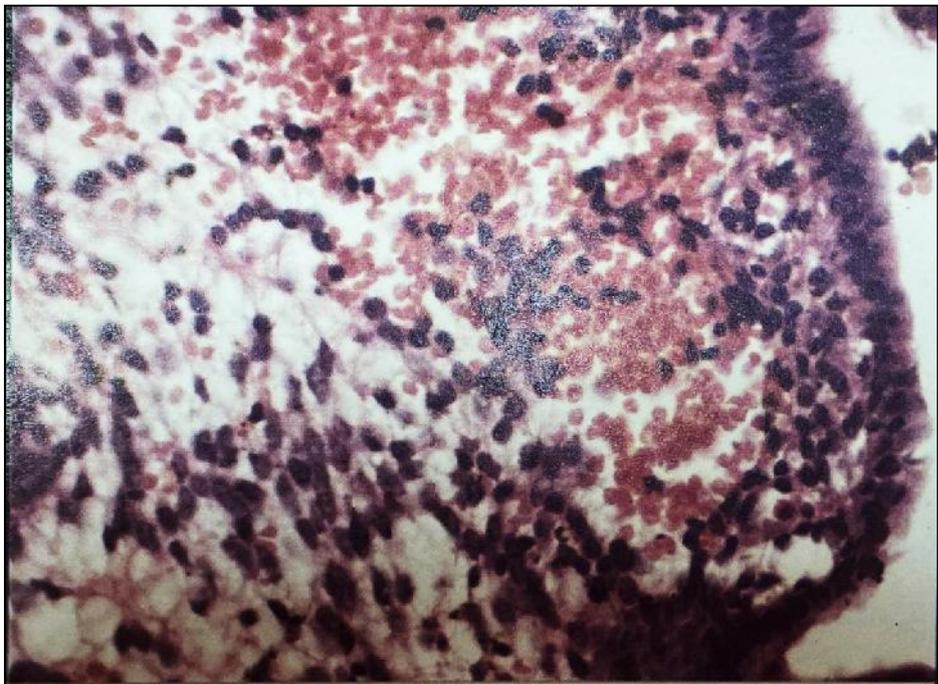
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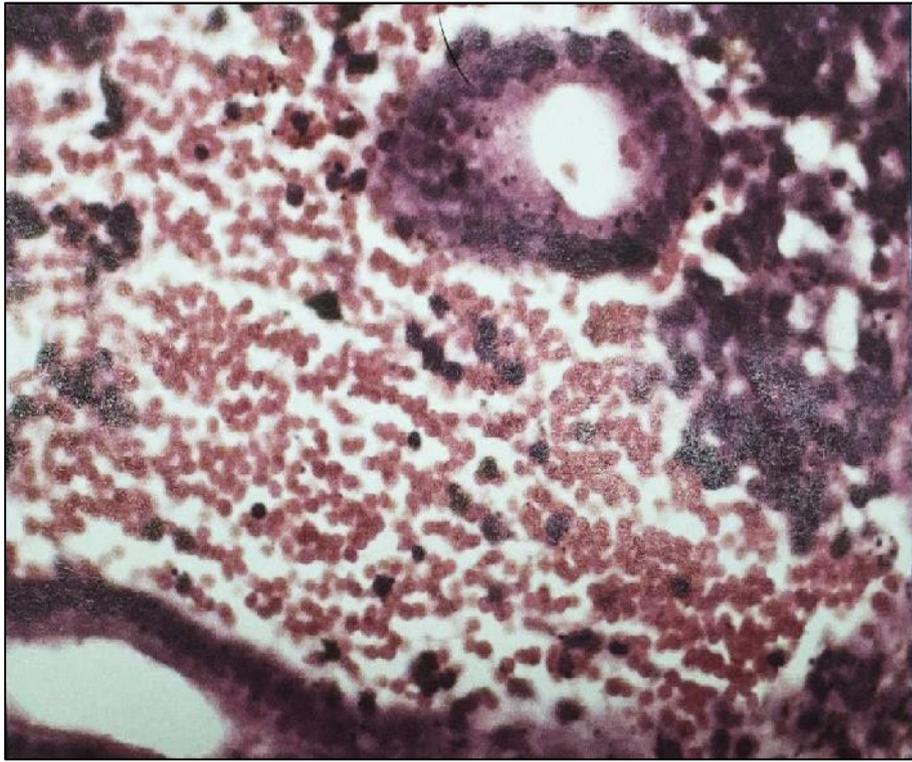
13 -

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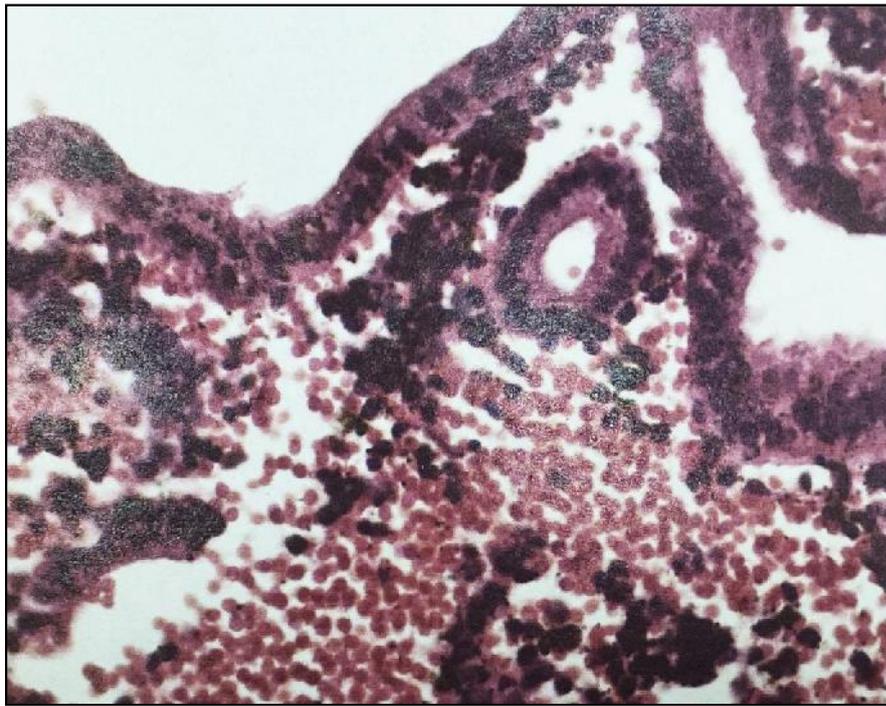
15 -

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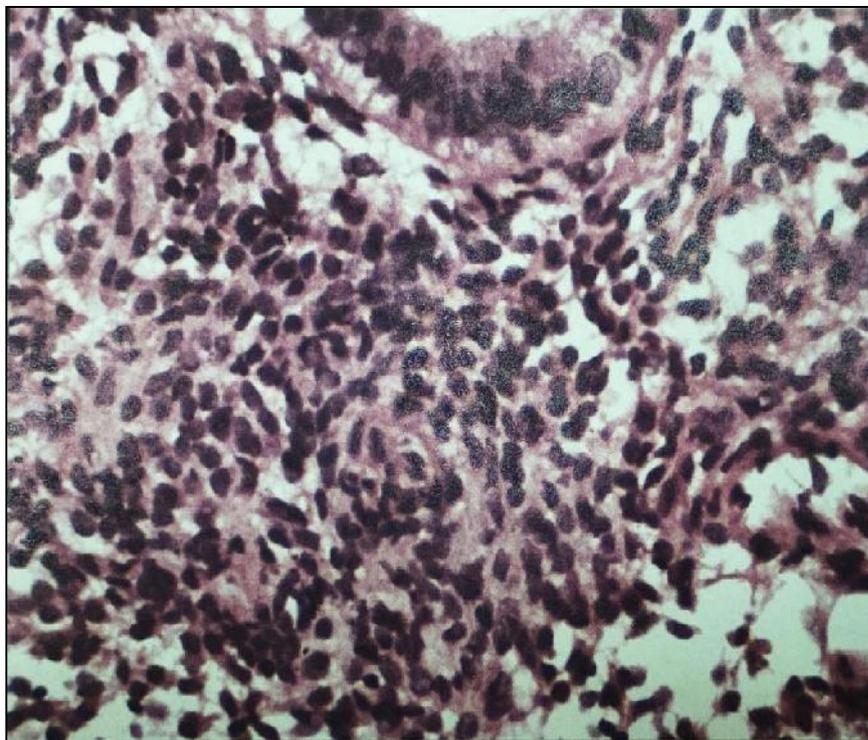
18, 19).

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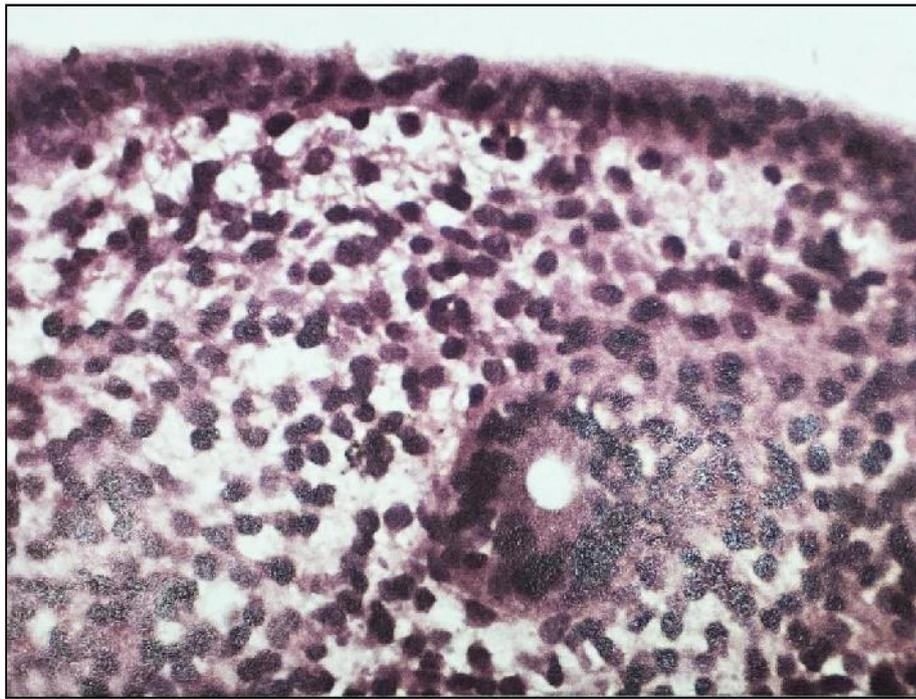
16 -

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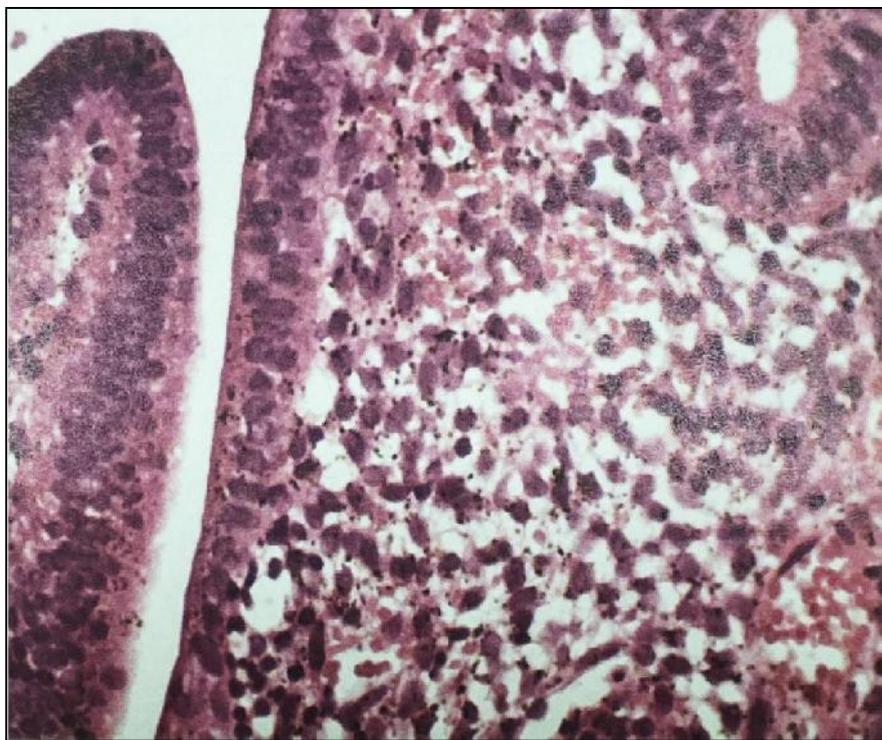
17 -

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×40



18 -

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×40



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×40

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 6 - VEGF, TGF-

	1- (n=44)		2- (n=42)	
	.		.	
VEGF	140 [100;184]	135 [100;215]	184 [172;280]	190 [170;220]
TGF-	130 [100;170]	135 [95;180]	105 [70;150]	100 [70;120]

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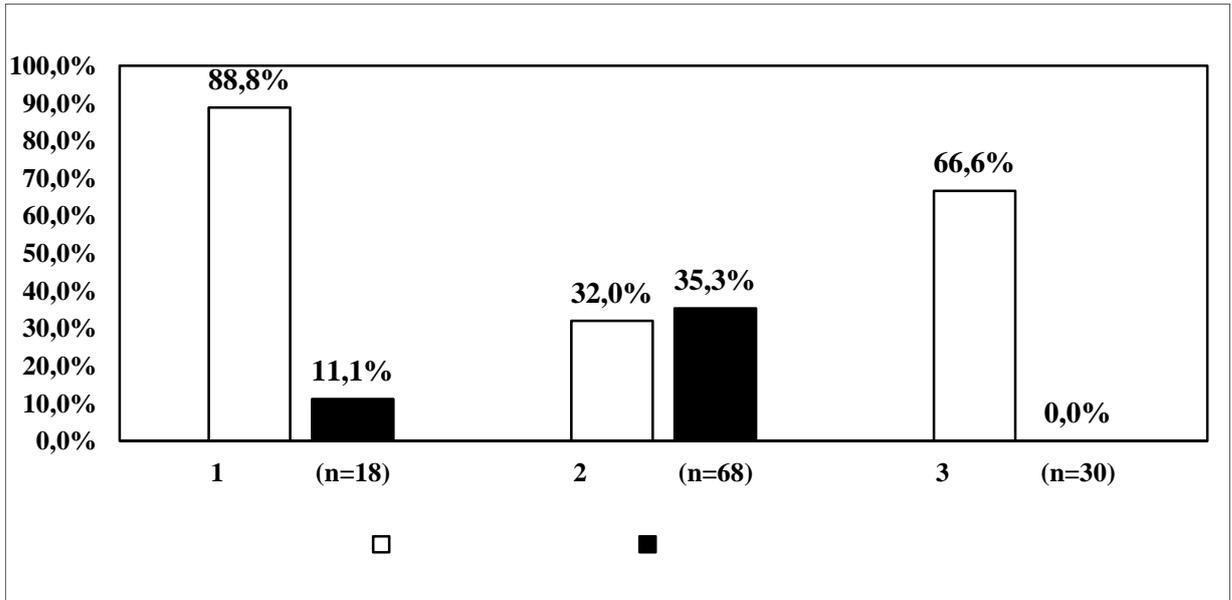
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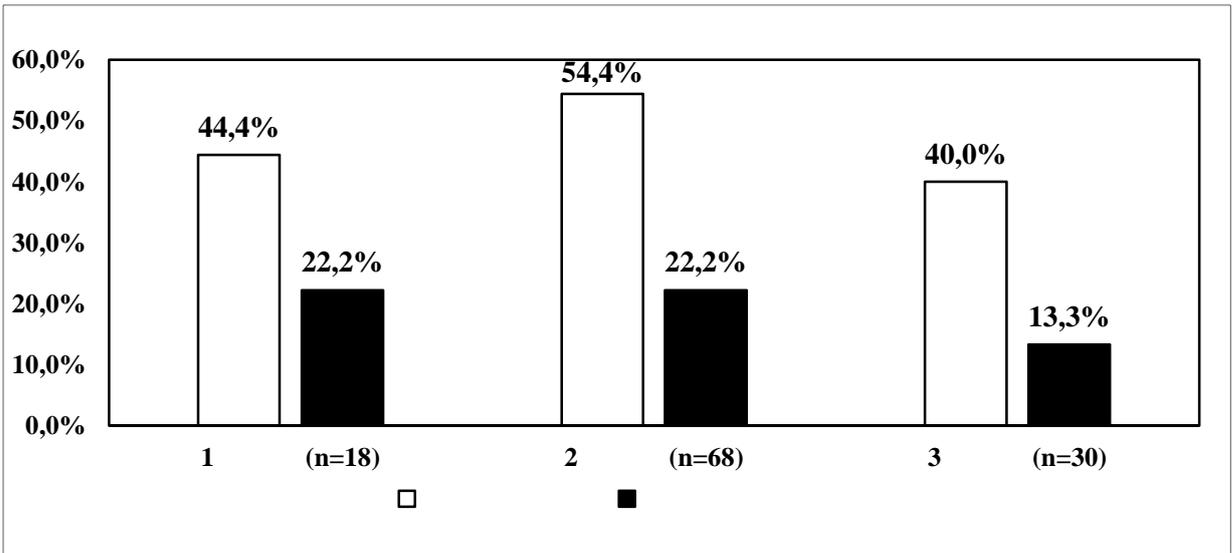
: 1- – 89%,
 66%
 2- –
 54,4%, 1- 44,4% , 3- – 40%

7 –

	1- (n=18)	2- (n=68)	3- (n=30)
	13 [12; 15]	13 [11; 15]	13 [12; 16]
	28 [24; 32]	27 [22; 34]	28 [25; 32]
	16 [88,8%]	32 [47%]	20 [66,6%]
	8 [44,4%]	37 [54,4%]	12 [40%]



20 –



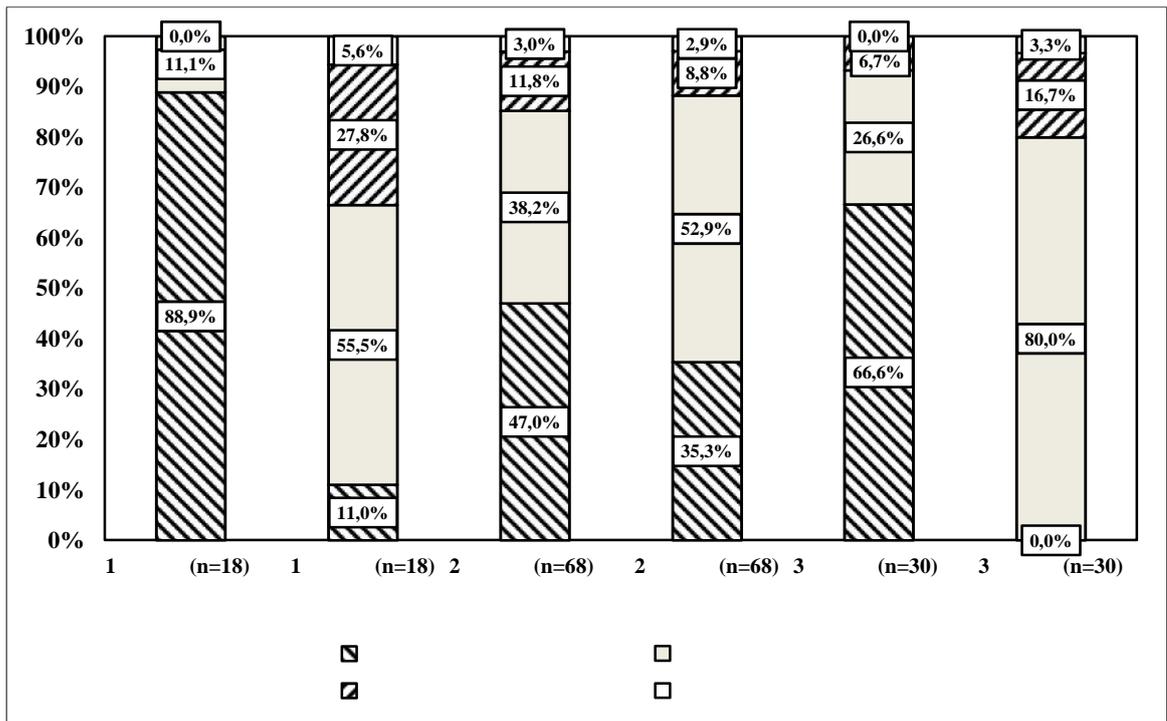
21 –

, (1-),
 2- , - 4-
 , 22,2%. -

13,3%

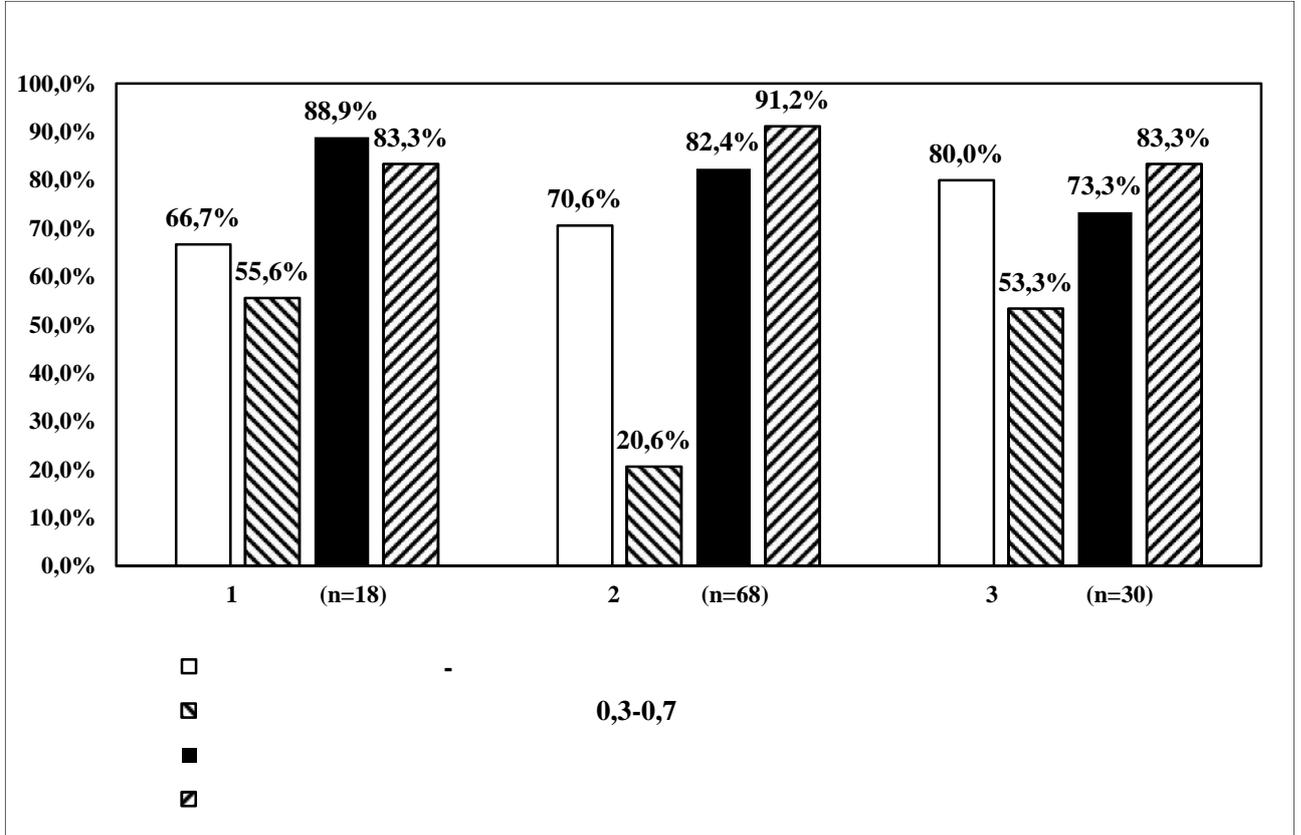
(22).

8.



) [123].

23.



23 –

(>0,05).

9.

– 3,4 , 3,5 , – 6,8 ; – 7,1 ; – 3,5

7,5 .

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10.

9 –

	1- (n=18)	2- (n=68)	3- (n=30)
,	3,5 [2; 7,8]	3,4 [2; 7]	3,5 [0,9; 6,4]

10 –

	1- (n=18)		2- (n=68)		3- (n=30)	
,	3,5 [2; 7,8]	7,1 [3,1; 9,1]	3,4 [2; 7]	6,8 [3,7; 7,0]	3,5 [0,9; 6,4]	7,5 [4,5; 8,0]

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«

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11.

11 –

()	-					
	1- (n=18)		2- (n=68)		3- (n=30)	
	IR	IR	IR	IR	IR	IR
	0,83 [0,74; 0,96]	0,79 [0,73; 0,87]	0,86 [0,76;0,98]	0,82 [0,75;0,90]	0,80 [0,65; 0,87]	0,78 [0,75; 0,87]
	0,80 [0,68; 0,90)	0,74 [0,61; 0,84]	0,80 [0,70;0,91]	0,80 [0,69;0,89]	0,80 [0,74; 0,89]	0,78 [0,65; 0,82]
	0,68 [0,54; 0,79]	0,64 [0,50; 0,72]	0,70 [0,60;0,80]	0,68 [0,60;0,78]	0,68 [0,56; 0,8]	0,71 [0,45; 0,80]
	0,69 [0,55; 0,95]	0,60 [0,45; 0,69]	0,65 [0,53;0,76]	0,60 [0,58;0,72]	0,65 [0,56; 0,78]	0,60 [0,54; 0,68]
	0,00 [0; 0,78]	0,51 [0,43; 0,62]	0,5 [0;0,67]	0,47 [0,30;0,59]	0,52 [0; 0,74]	0,45 [0,30; 0,56]
	0 [0; 0]	0,5 [0,39; 0,58]	0 [0;0]	0,45 [0,35;0,53]	0 [0; 0]	0,42 [0,38; 0,45]

(>0,05).

5.3

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7-8-

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3-4-

24.

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25.

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(>0,05).

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8.

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3,5 ,

-7,1 ;

-3,4 ,

-6,8 ;

-3,5

-7,5

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,

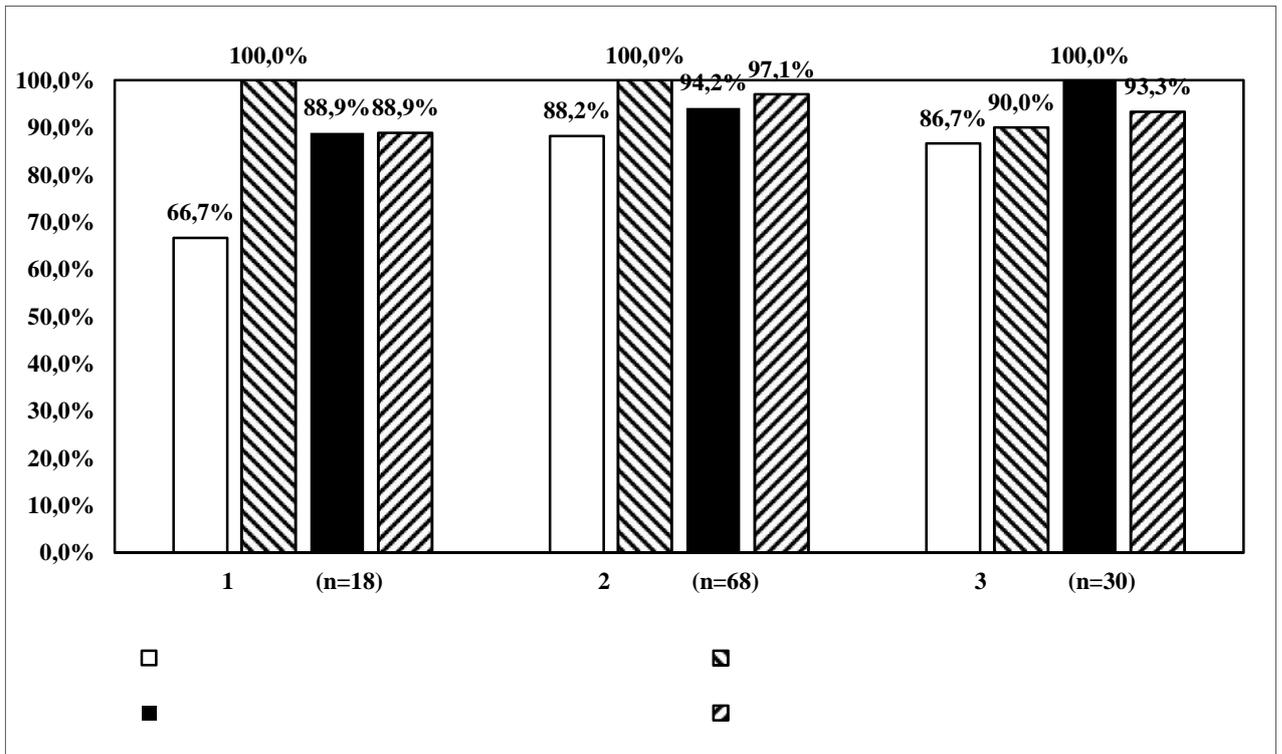
,

-

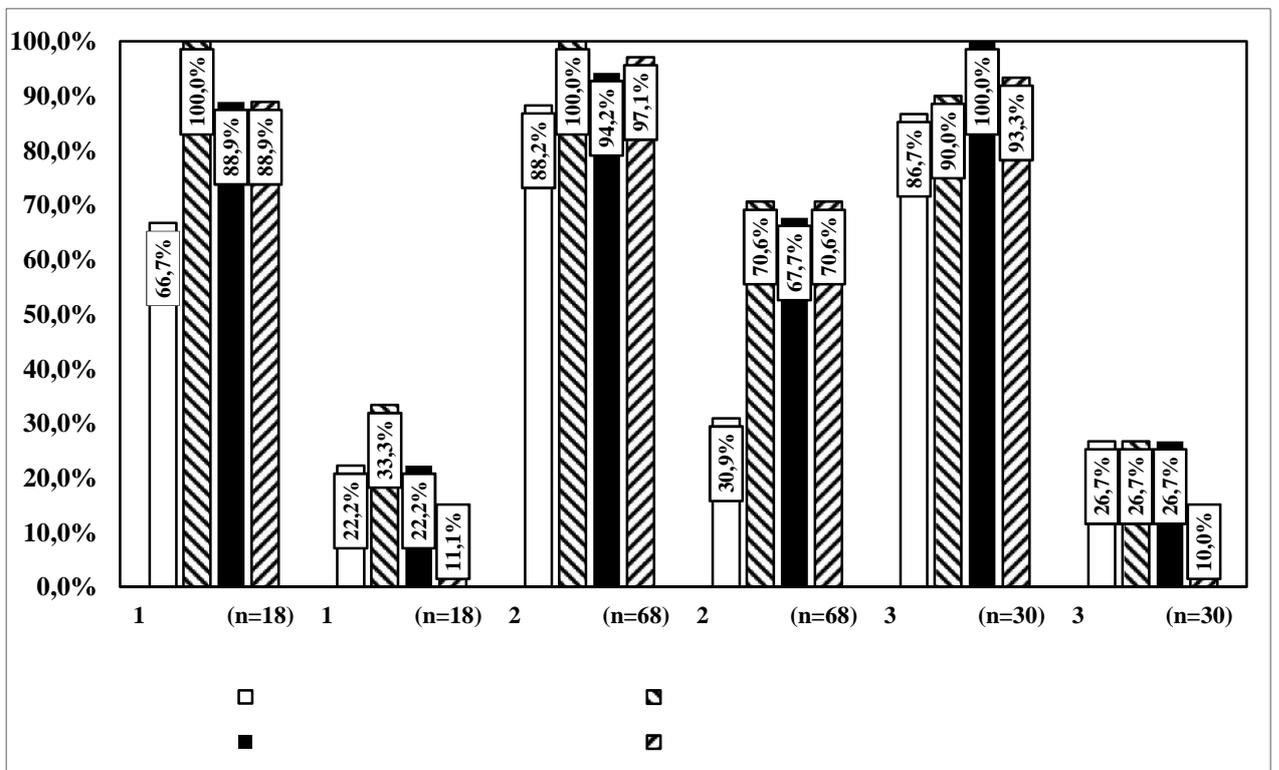
-

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10.

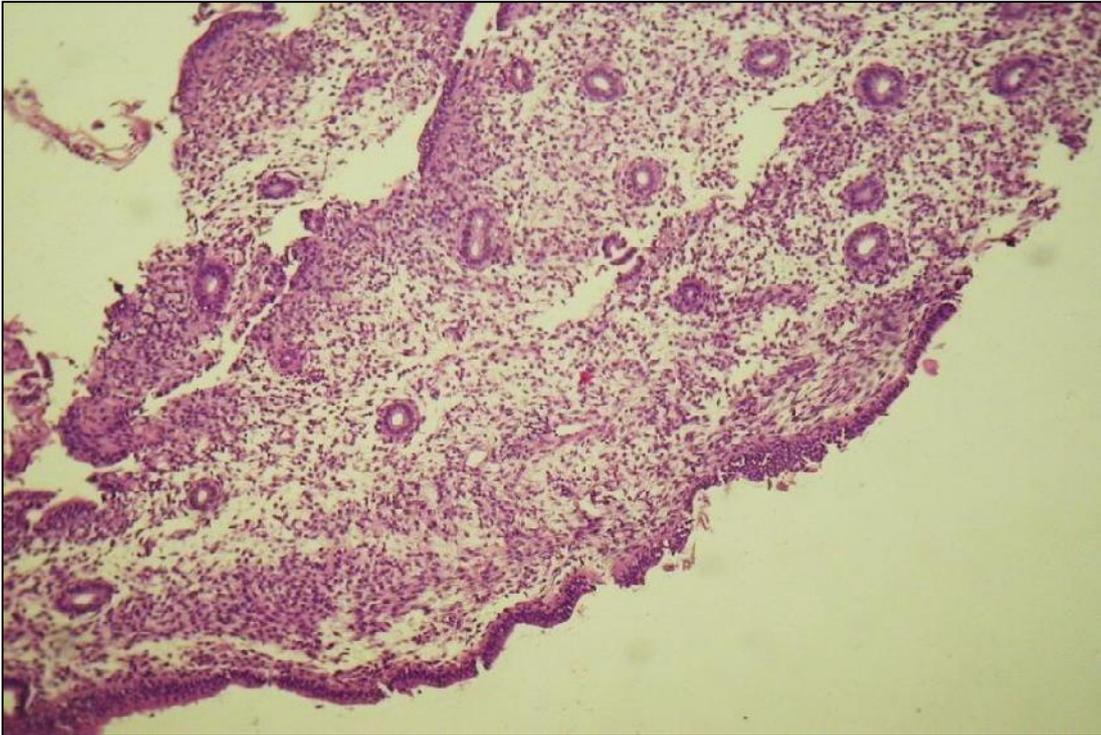


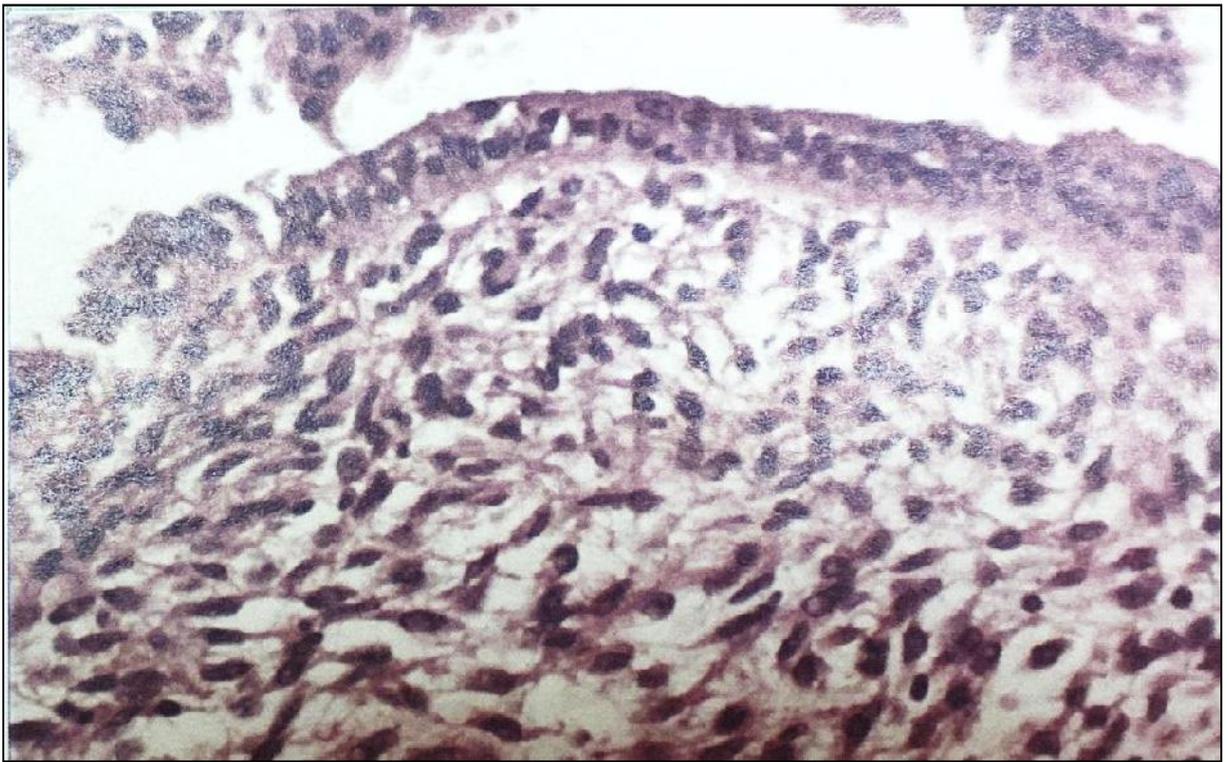
24 –



25 –

71
 . 1- -
 4- (22,2%), -
 6 (33,3%), - 4 (22,2%),
 2 (11,1%) . -
 21 (30,9%) , -
 48 (70,5%) , -
 46 (67,4%) , 48 (70,5%) .
 8 (26,6%), 3-
 (10%) (26).



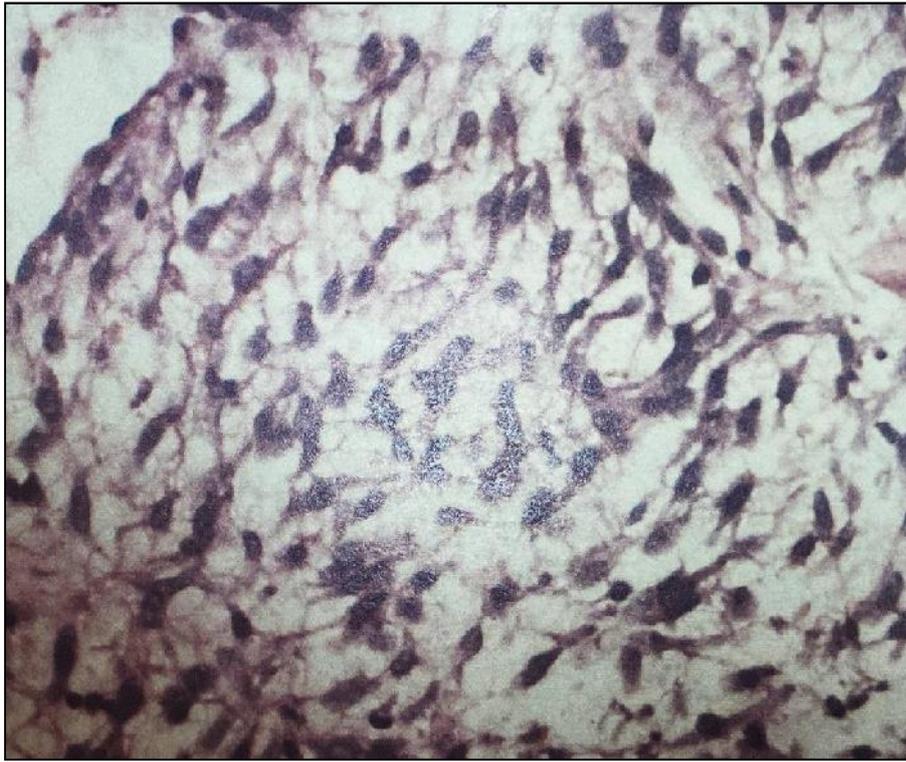


28 -

:

×40

(29, 30).

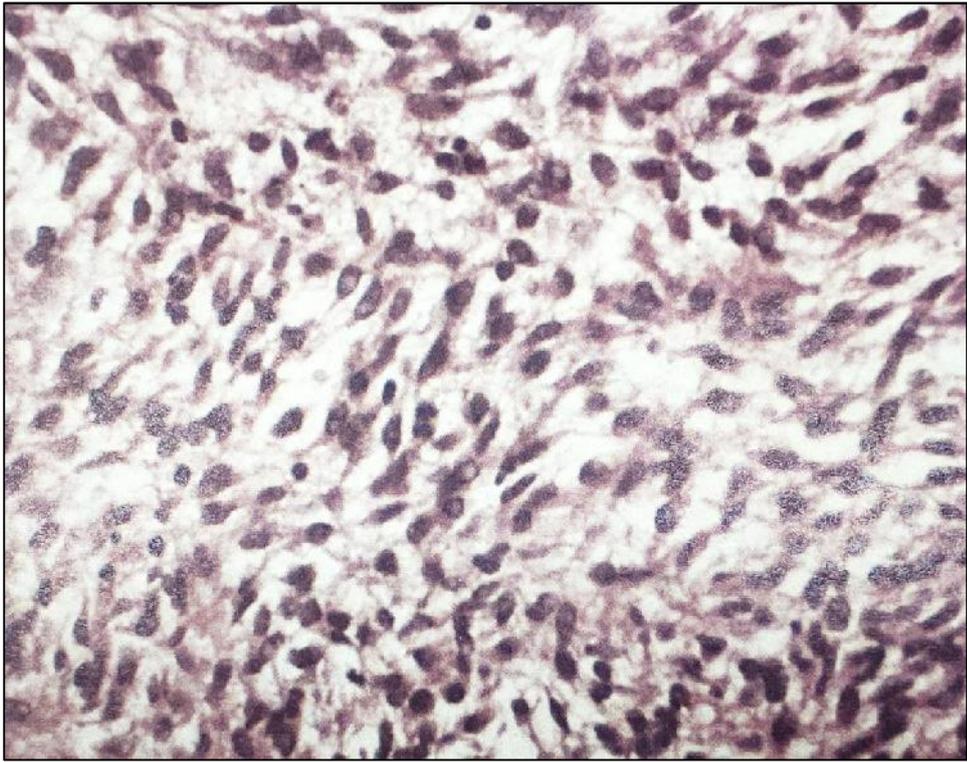


29 –

×40

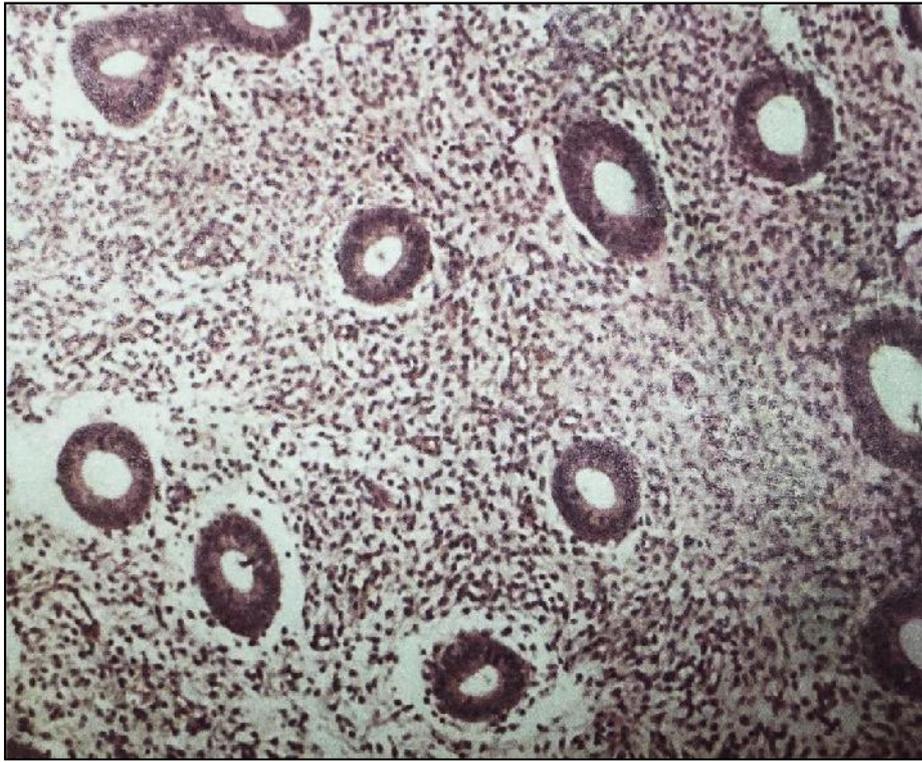
()

(31, 32).



30 –

×40

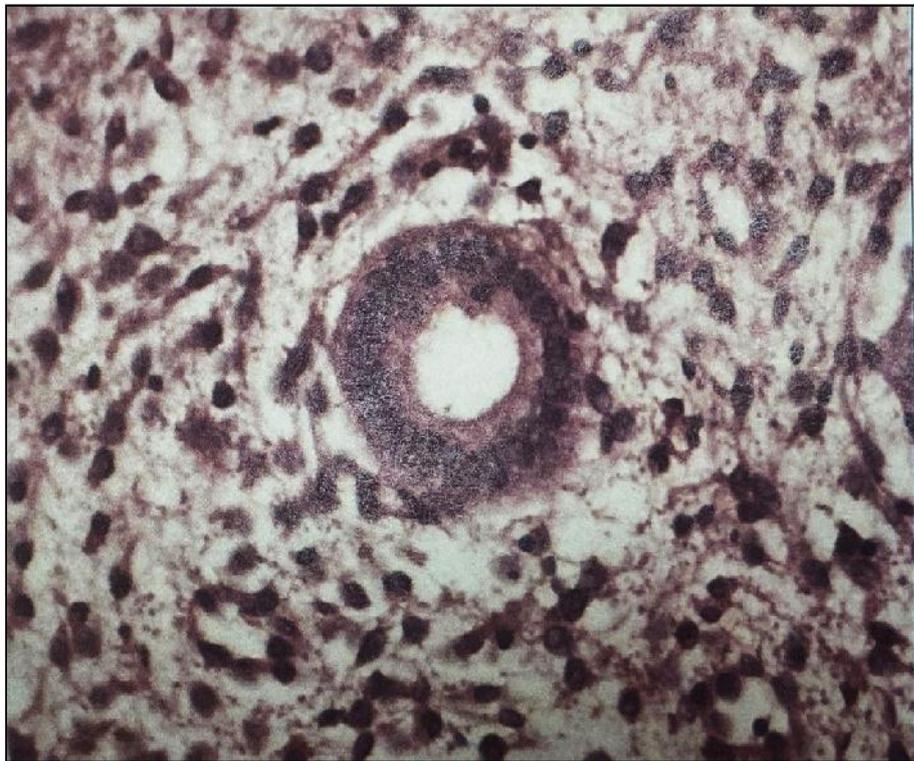


31 -

,

,

×40



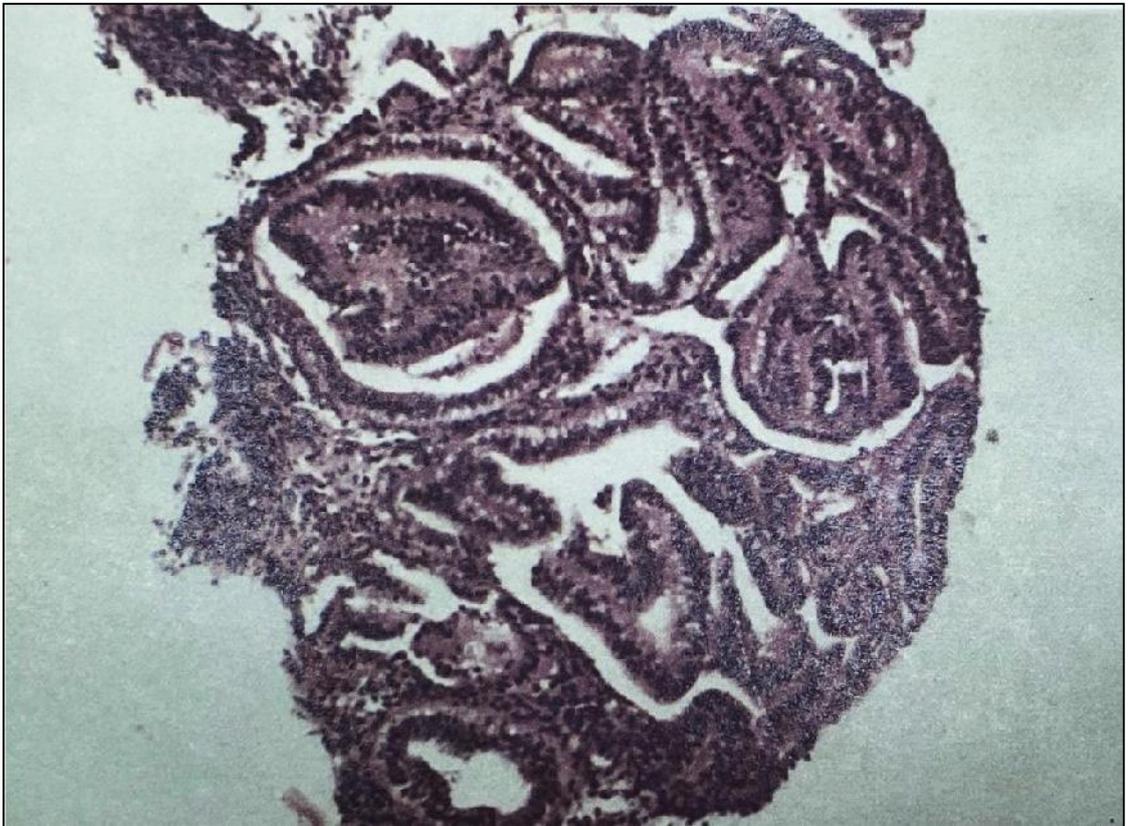
32 -

()

,

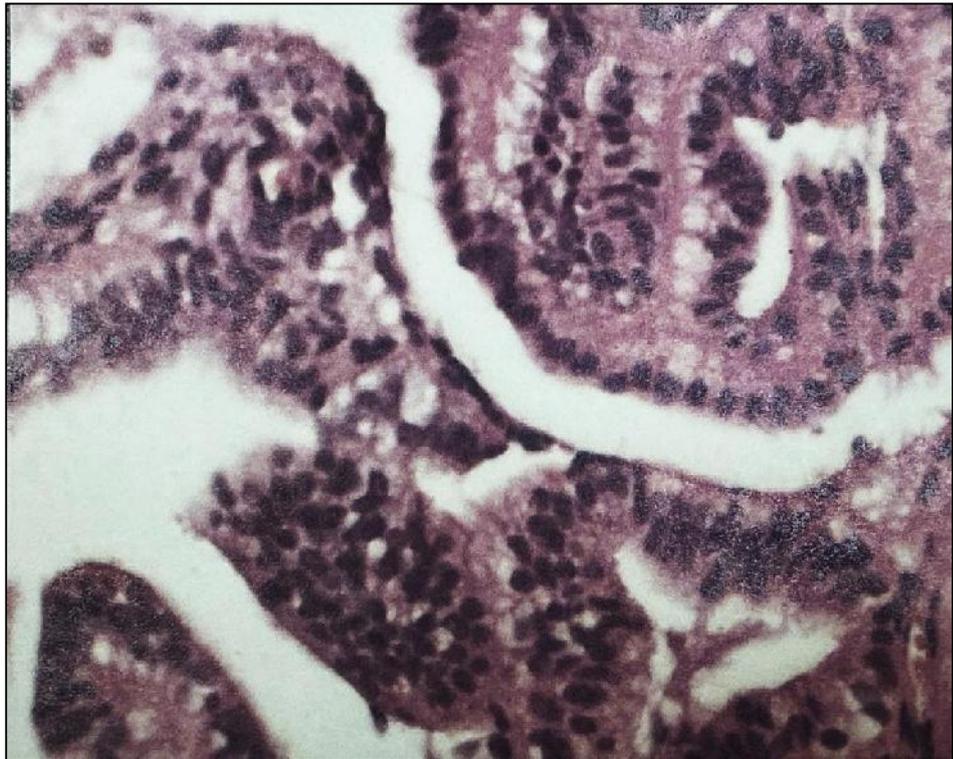
×40

(33, 34).



33 –

×40



34 -

×40

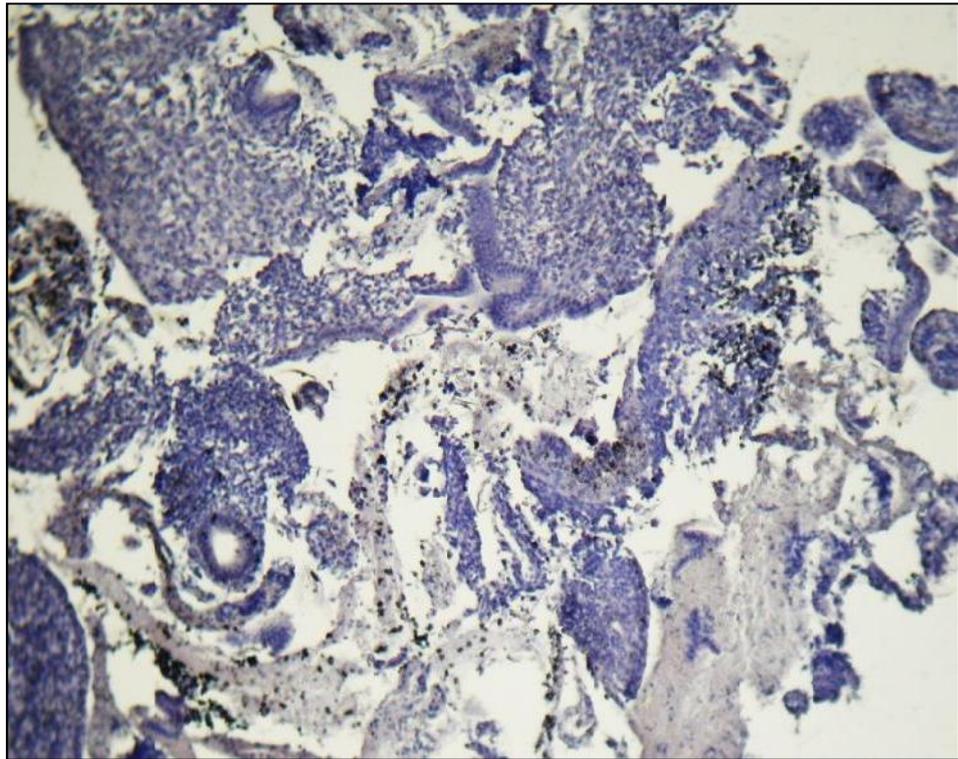
CD138 48
 ; CD138
 (36).

VEGF Hscore (12).

, VEGF
 145 (100; 195), - 130 (100; 205);
 - 145 (95; 200), - 125 (90; 170);
 150 (100; 210), - 130 (70; 200),
 (37).

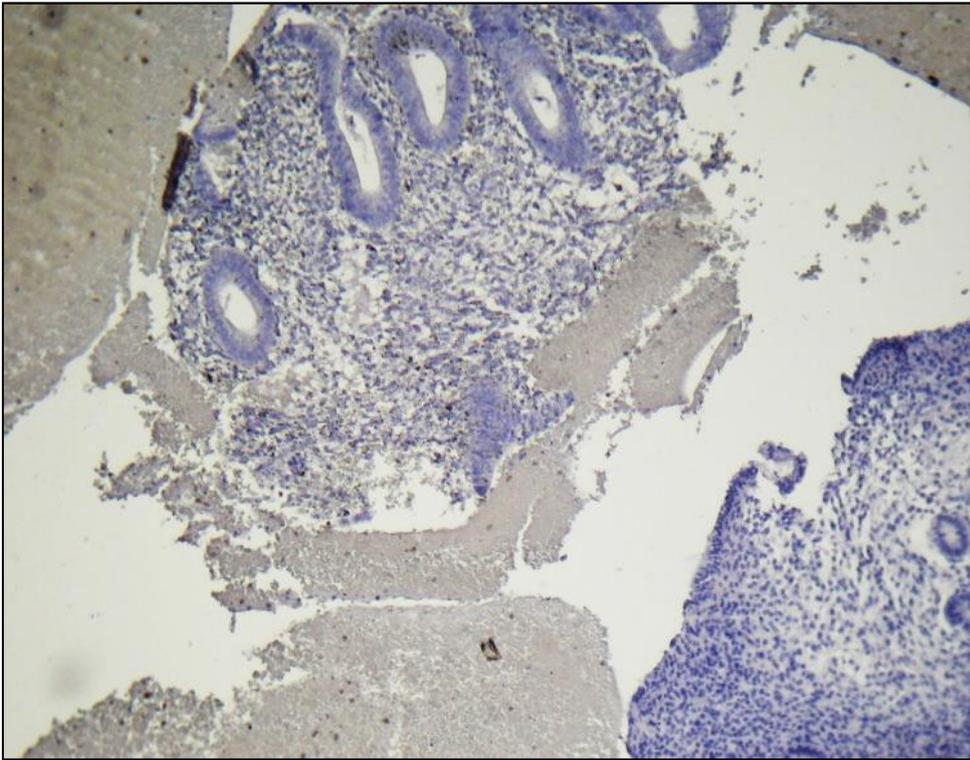
TGF-

(38).



35 – CD138

10x. 10x



36 –

CD138

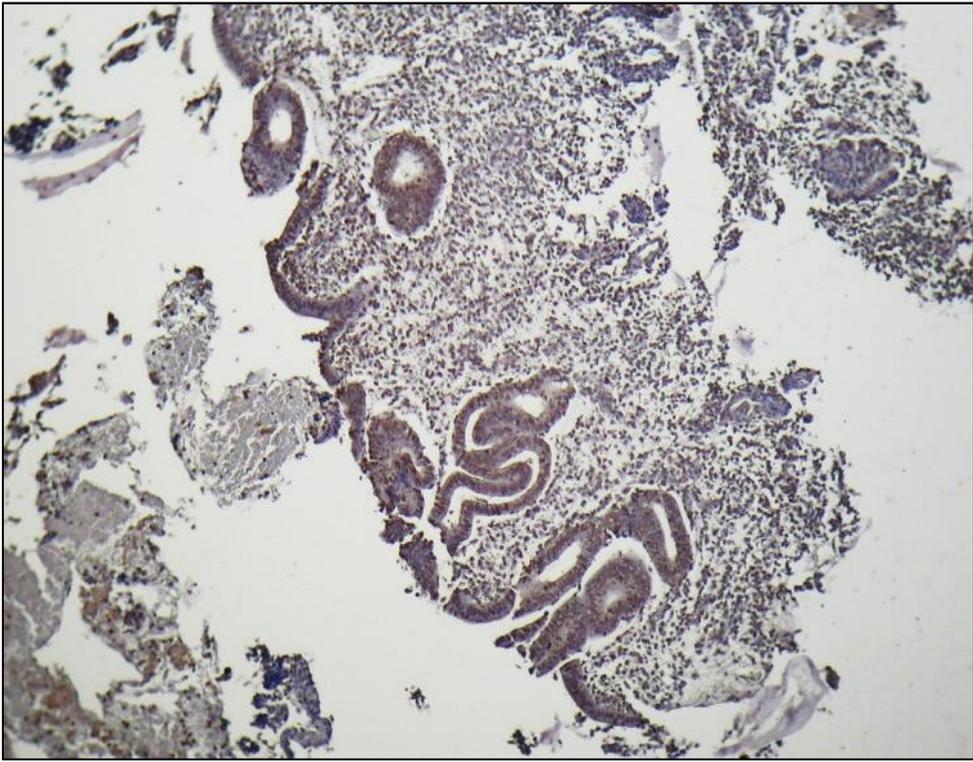
10×

10×

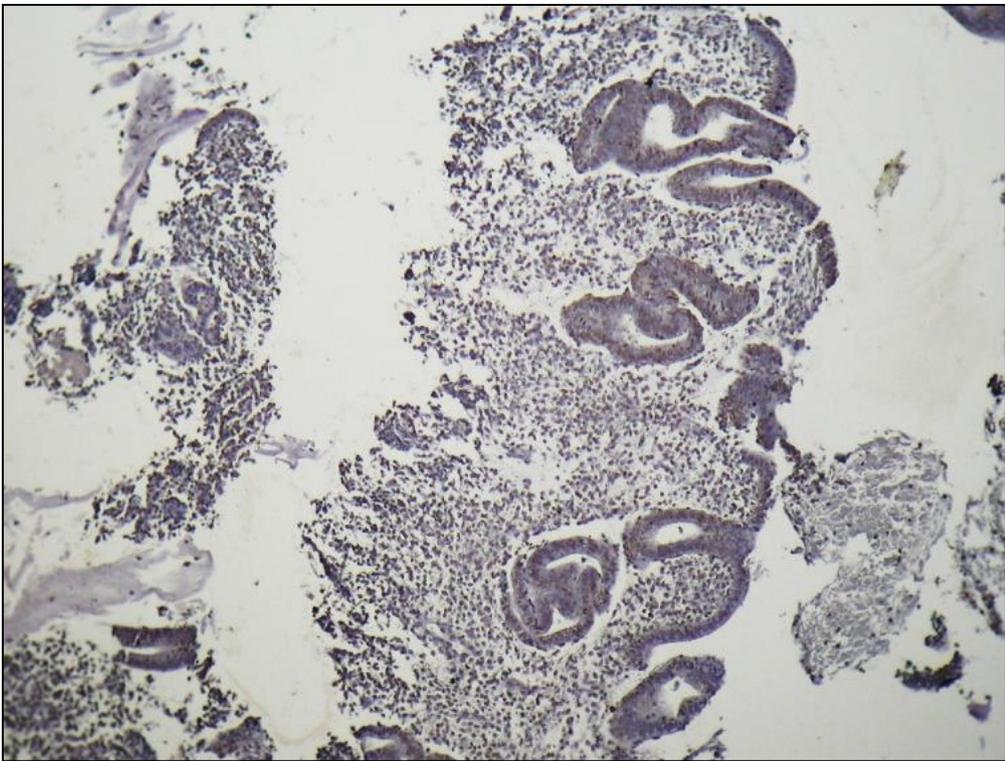
12 –

VEGF, TGF-

	1- (n=18)		2- (n=68)		3- (n=30)	
	.		.		.	
VEGF	145 (100;195)	130 (100;205)	145 (95;200)	125 (90;170)	150 (100;210)	130 (70;200)
TGF-	130 (100;170)	135 (95;180)	130 (100;170)	130 (100;180)	125 (95;180)	130 (100;170)

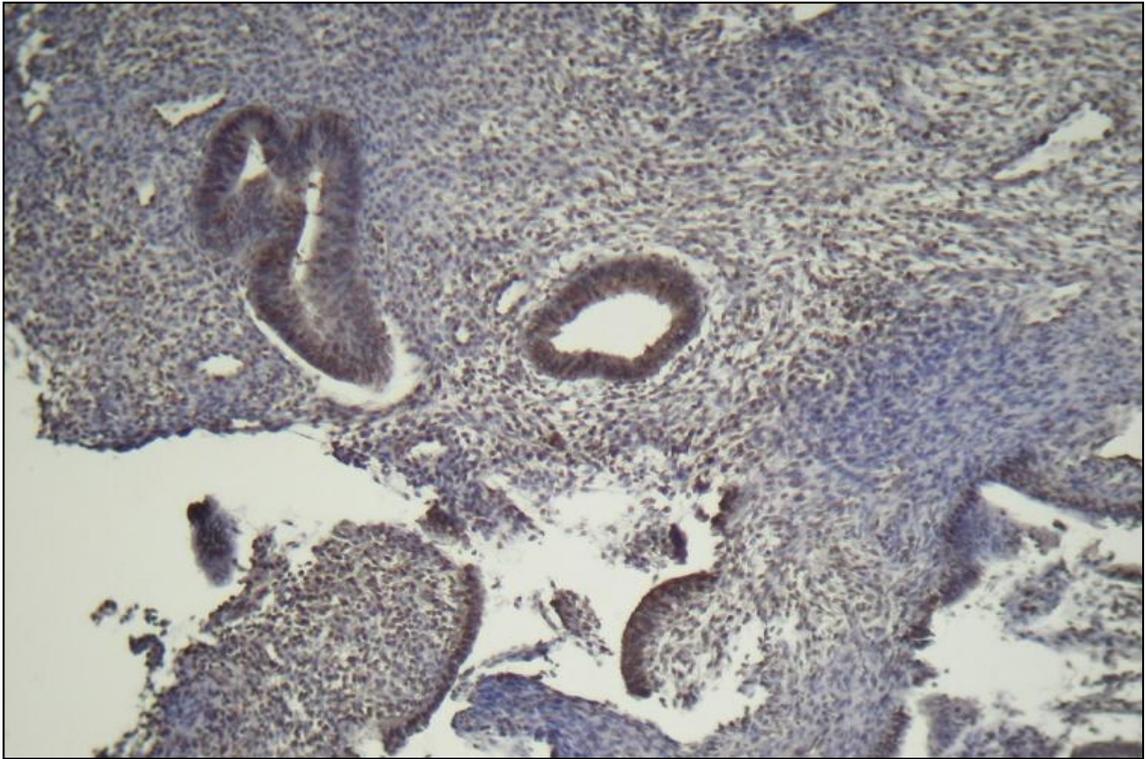


37 – VEGF . . .
10x. 10x



38 – TGF- . . .
10x. 10x

220 (180; 240), 20 (170; 240) (39).



39 –

VEGF

10x.

10x

190 (180; 200),

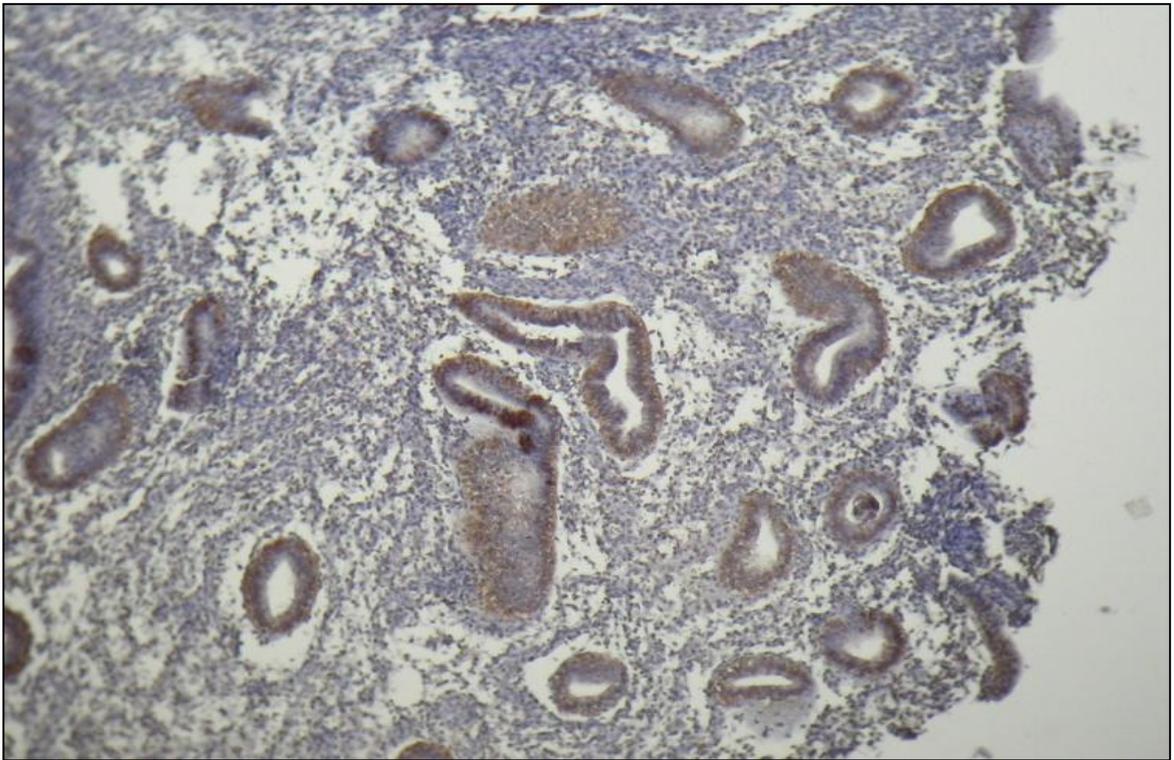
– 180 (170; 210).

VEGF

180 (170; 200),

– 185 (170; 210),

(70; 150) , TGF- 130 (100; 170) 120 -
 : - 135 (95; 180), - 100 (75; 130).
 TGF- ,
 , 110 (65; 145) (40). -
 VEGF, TGF- -
 41, 42.

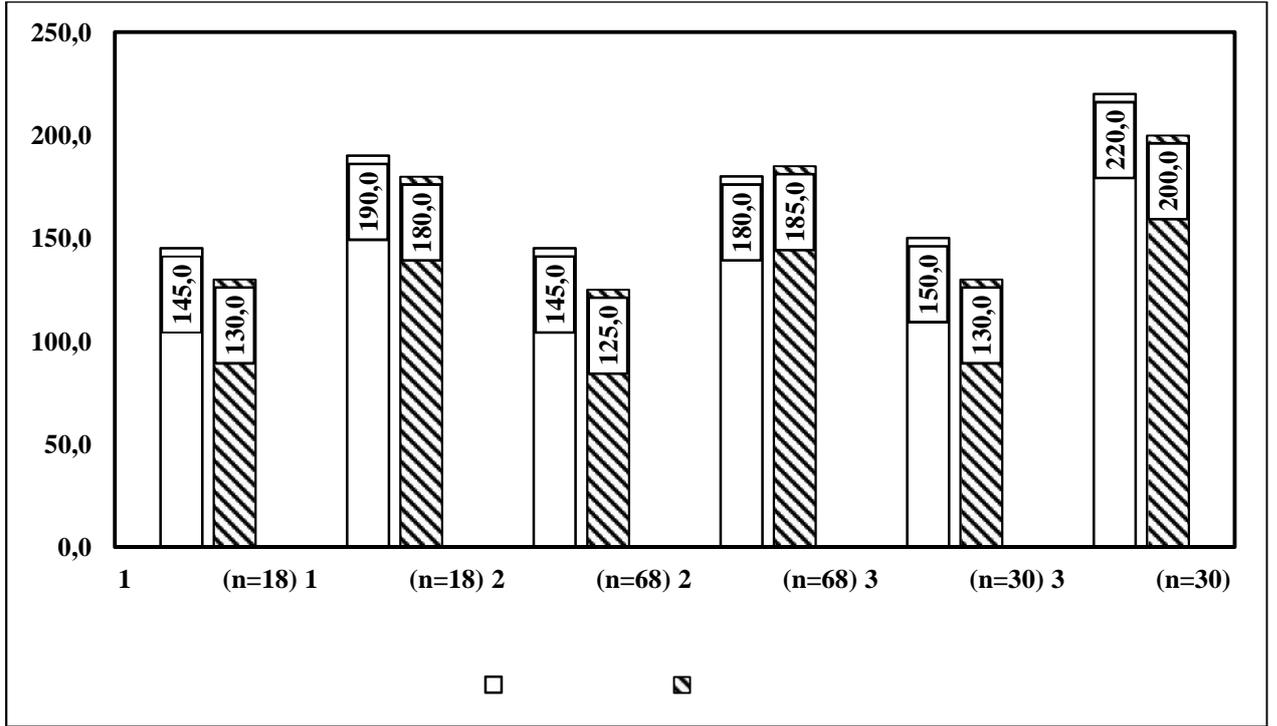


40 -

TGF-

10x.

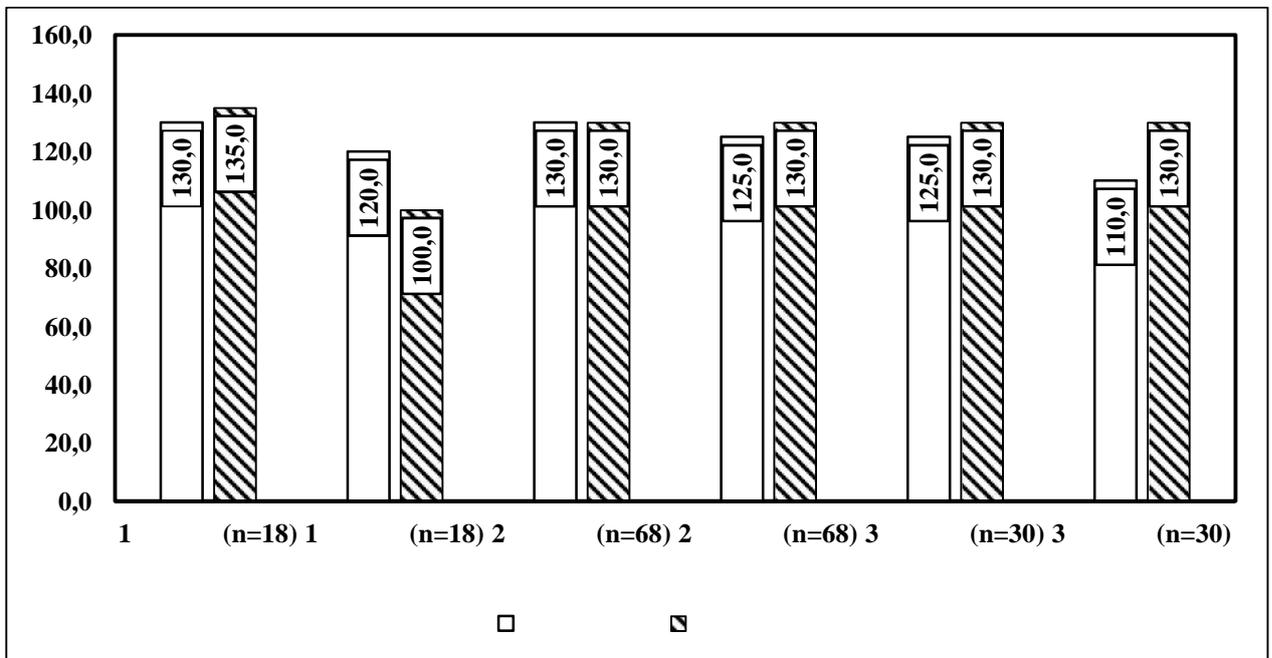
10x



41 -

VEGF

(Hscore)



42 -

TGF-

(Hscore)

6

(71 – 2010 2015 . 140 – 2015 2020 .)
(PubMed, 2020). , -
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,
(RIF). -
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() CD 138
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-
(Kannar V., Lingaiah H.K., Sunita V., 2012; Buzzaccarini G. et al., 2020).
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,
(-
. , 2018).
-
1
– TGF- 1. TGF-b1 / Smad
(Liu L. et al., 2019;
Wei Z. et al., 2020). -

- (-) 8

(. . . , 2019; . . . , 2019; . . . , 2020; . . . , 2020).

, (Maged A.M. et al., 2019; Akgün Kavurmacı S. et al., 2021).

(VEGF).

VEGF

86

(48% 1- , 46% 2-)

1- 2-

(

88

):

– 80,9%

– 50%.

, 2020).

(52%)

(86%).

(46%),

5,4 (3,8; 7,7)

3,2 (2,3; 6,8)

0,47 (0,20; 0,60).

3–4

(81,81%),

(28,57%).

CD

138

VEGF

TGF-

VEGF

TGF-b.

,
 ,
 (Tandulwadkar S.R. et al., 2017). PRP –
 « »
 ,
 (Sfakianoudis K. et al., 2019) (Galin-
 do M.P. et al., 2016; Maleki-Hajiagha A. et al., 2020; Tsai W.C. et al., 2021).

,
 (. . . , 2019;
 . . . , 2020).

(Duan Z. et al., 2021).

22,2%.

8-

13,3%

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 .
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 -
 ,
 ,
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 - - .
 -
 . «
 »
 - :
 - 3,5 (2; 7,8) , - 7,1 (3,1; 9,1) ,
 - 3,4 (2; 7) , - 6,8 (3,9; 7,8) ,
 - 3,5 (0,9; 6,4) - 7,5 (4,5; 8,0) .
 .
 100%
 « ».
 PRP- IR
 , (0,45 (0,35; 0,53))
 (0,42 (0,38; 0,45)) ,
 .
 3-
 . 3-
 .

91

3-4 (

CD 138-

CD 138

CD138

4 (22,2%),

6 (33,3%),

2 (11,1%)

48 (70,5%)

46 (67,4%)

7 (23,3%),

3- (10%).

PRP-

VEGF TGF-

70

VEGF Hscore

VEGF

:

- 45/35

, - 50/60 .

VEGF PRP- ,

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, PRP-

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TGF-

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, PRP-

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1. , -
, VEGF,
TGF- .
2. -
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3. .
TGF- , -
4. , -
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VEGF.

1. , -
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EFG –

EGF –

FGF –

GF –

L F –

PDGF –

PRP – ,

TGF –

Th 1,2 – - 1,2

TNF –

VEGF –

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1. . . . , . . . , . . . , . . . // -
 . - 2016. - 4-2 (96). - . 41-46.
2. . / . . . ,
 . . . , . . . , - ∴ - ,
 2014. - 1200 .
3.
 / . . . , . . . , . . . , . . . // -
 . - 2016. - 3. - . 336-341.
4. -
 ∴
 / . . . , . . . , - ∴ Status
 Praesens, 2013. - 16 c.
5. , . . .
 / . . . ,
 . . . , . . . // . - 2012. - 2
 (12). - . 27-30.
6. , O.A. -
 ∴
 / O.A. , . . . //
 . - 2012. - 4. - . 27-30.
7. ∴ / -
 ∴ - , 2020. - 432 .
8. , . . . -
 /

- ... , ... , ... // ... -

2017. – 2. – .90–96.

9. , ... -

/ ... ,

... // ... – 2018. – .24, 6. – .67–75.
- 10. , ... -

/ ... ,

... // ... – 2016. – 1–2. –

.13–16.
- 11. , ... « » -

– – /

... , ... , ... //

... – 2014. – 7 (39). – .11.
- 12. , -

« » / ... , ... , ... -

... // ... – 2016. – .3, 27 (290). –

.39–43.
- 13. :

/ ... , ... ,

... [.] // ... , -

... – 2021. – .20, 5. – .43–50.
- 14. , ... « »:

... : 14.01.01 / ... , 2014. – 22 .
- 15. . / ... , ... -

... [.]. – .: – , 2020. – 544 .

16. , -
 : KI-67, P16INK4A
 D (VDR) / . . , . . , . . ,
 . . // . - 2019. - 5 (173). -
 . 87-94.
17. , -
 : - / . . , . . -
 , . . , . . // -
 . - 2020. - 3 (186). - . 36-41.
18. , . . -
 / . . , . . , . . //
 - . - 2015. - . 15, 1. - . 32-37.
19. , . . -
 / . . , . . //
 . - 2015. - 11. - . 78-85.
20. , . . -
 () / . . , . . //
 . - 2012. - 9. - . 42-46.
21. , . . -
 / . . , . . //
 . : , . - 2013. - . 11,
 2. - . 99-104.
22. / .
 : <https://minzdrav.gov.ru/opendata/7707778246-grls> (-
 01.02.2018)
23. , . . -
 , -

- / . . . , . . . , . . . // -
 , . - 2013. - 3. - . 35-50.
24. , . . . /
 . . . // . - 2013. - 4 (92). -
 . 18-20.
25. (Divigel®) / VIDAL: -
 :
www.vidal.ru/drugs/divigel_1125 (: 01.03. 2018).
26. , . . . -
 /
 . . . , . . . // . - 2019. - 9.
 - . 139-146.
27. , . . . ? / . . . // Status Praesens. - 2021.
 - 4 (79). - . 17-23.
28. : . RU 176780 U1 . /
 . . ;
 " - ". - 2017118118; . 24.05.2017;
 . 29.01.2018.
29. , . . . :
 . : 14.01.01 / . - ., 2012. - 24 .
30. - -
 / . . . , . . . ,
 . . . [.] // .
 - 2013. - . 76, 3. - . 27-30.

31. — /
 . . . , . . . , . . . [.] //
 . – 2019. – 12. – . 154–160.
32. - /
 . . . , . . . , . . . , . . . //
 . – 2017. – 1, 3 (300). – . 46–50.
33. - /
 " " / . . ,
 . . . , . . . , . . . // -
 . – 2016. – 22, 3. – . 57–62.
34. / . . ,
 . . . [.] // -
 – 2012. – 6. – . 95–101.
35. - / . . , . . ,
 . . // -
 . – 2013. –
 1. – . 155–158.
36. - / . . , . . , . . //
 . – 2017. – 3. – . 29–32.
37. -
 -

- / . . . , . . . [.] // -

. - 2015. - . 17, 3. - . 93-96.
- 38. , . . . -

() / . . . , . . . // -

. - 2013. - 4. - . 21-26.
- 39. , . . . -
- / . . . // -

. - 2021. - . 27, 4. - . 117-124.
- 40. , . . . -

() / . . . , . . . -

, . . . // . - 2016. - . 22, 5. -

. 61-69.
- 41. , . . . « » -2

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- / . . . , . . . // -

. - 2017. - . 66, 4. -C. 46-50.
- 42. , . . . -

/ . . . , . . . // . - 2012. -

3. - . 49-53.
- 43. , . . . -

/ . . . , . . . , . . . // -

. - 2018. - . 16, 6. - . 99-105.
- 44. -

: . 2747943 C2 . / . . ,

. – 2018113907; . 17.04.2018; .

17.05.2021.

45. , . . . -

/ . . . -

, . . . , . . . // .

– 2017. – 9. – . 119–127.

46. -

/ . . . , . . . , . . .

[.] // . – 2017. – . 6, 51. – . 80–83.

47. -

/ . . . , . . . -

, . . . [.] // , -

. – 2018. – . 17, 1. – . 27–32.

48. -

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. . . , . . . , . . . [.] //

. – 2019. – 5. – . 125–132.

49. : -

: -

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[.] – . : StatusPraesens, 2021. – 68 .

50. « »

/ . . . , . . . , . . .

[.] // VIII

. – ., 2014. – . 262–264.

51. -

« » / . . . ,

. . . , . . . [.] // . –

2021. – 4. – . 112–119.

52. , . . . () /
. . // . – 2013. – 1. –
. 224.
53. / . . , . . , . . ,
. . // – . – 2017. –
. 17, 5. – . 29–34.
54. , . . : , , -
/ . . , . . , . . -
// . – 2020. – . 9, 2. –
. 16–25.
55. , . . – , -
/ . . , . . //
. – 2018. – . 67, 1. – . 38–46.
56. / . . , . . , . . [.] //
. – 2019. – 8. – . 100–106.
57. , -
/ . . , . . , . . [.] //
Journal of Siberian Medical Sciences. – 2019. – 1. – . 28–35.
58. , . . / . . ,
. . // . – 2014. – . 9, 2. – . 65–73.
59. , -
,

/ . . . , . . . [.] //
. – 2020. – 4. – . 82–89.

60. , . . . -
/ . . . , . . . , . . . //
. – 2018. – 6. – . 10.

61. , . . . -
/ . . . // . –
2016. – 3. – . 113–118.

62. , . . . -
/ . . . ,
. . . , . . . // -
. – 2014. – 2. – . 57–64.

63. , . . . //
/ . . . , . . . //
. – 2013. – 1. – . 44–53.

64. -
/ . . . ,
. . . , . . . [.] // ,
. – 2017. – . 16, 6. – C. 36–43.

65. . -
().
2.0 / [.] – .: StatusPraesens, 2020. – 128 .

66. () -
() / . . . , . . . ,
. . . [.] // . . . -
. – 2015. – . 2, 4. – . 51.

67. -

- / . . . , . . . , . . . [.] // -
 , - 2019. - . 18, 6. - . 34-40.
68. / . . . , . . . [.] // . -
 2019. - 10. - . 172-179.
69. : -
 / . . . , . . . , . . . [.]. - .,
 2015. - 24 .
70. « » : -
 / . . . , . . . , . . . -
 [.] // . - 2020. - . 18, 8-9. - . 13-19.
71. , . . . / . . . -
 , . . . , . . . // . -
 2012. - 1. - . 72-76.
72. " " / . . . -
 , . . . , . . . [.] // . - 2017.
 - . 1, 12 (309). - . 34-38.
73. , . . . / . . . ,
 - ∴ - , 2013. - 304 .
74. / . . . , . . . ,
 . . . [.] // . . . -
 . - 2014. - . 1, 2. - . 48-49.
75. , . . . , -
 , (-

) / . . . , . . . , . . . // -
. - 2020. - . 26, 5. - . 91-98.

76. , . . . -
/ . . . , . . . //
. - 2018. - . 20, 6. - . 53-59.

77. , . . . : /
. . . , - ∴ - , 2014. - 1011 .

78. , . . . -
/ - ∴ -
, 2011. - 224 .

79.
/ . . . , . . . , . . . [.] //
- . - 2017. - . 17, 4. - C. 25-32.

80. -
/ . . . , . . . / , . . . ,
. . . // . - 2019. - . 68,
6. - C. 99-106.

81. « » :
2748490 / . . . , . . . , . . . -
. . . , -

26.05.2021.

82. , . . . : / . . . ,
. . . . - ∴ - , 2013. - 64 .

83. , . . . -
/ . . . ,
. . . , . . . // . - 2014. - 1. - . 104-109.

84. « » – /
 . . . , . . . , . . . [.] // . – 2018.
 – 6. – . 15–22.
85. , . . . -
 / . . . , . . . //
 – 2016. – . 3, 3. – . 141–148.
86. , . . . -
 / . . . , . . . // -
 . – 2012. – 1. –
 . 10–13.
87. , . . . -
 - : / . . . , . . . //
 . – 2019. – 5. – . 12–18.
88. -
 / . . . , . . . , . . . [.] // -
 . – 2020. – . 22, 3. – . 15–20.
89. : /
 . . . , . . . , . . . , . . . // -
 . – 2019. – . 21, 5. – . 49–52.
90. : , , ,
 / . . . , . . . , . . . //
 – . – 2013. – 5. – . 21–27.
91. , . . . -
 () // . . . (, . . .) // -
 . – 2012. – 3. – . 50–52.
92. , . . .
 / . . . // . – 2014. – 4.
 – . 4–6.

93. , . . .
 / . . . // Consilium Medicum. – 2013. – . 15, 6.
 – . 40-42.
94. P16INK4A , -
 / . . . , . . . -
 , . . . [.] // . – 2018.
 – 2 (157). – . 97–100.
95. A
 / . . . ,
 . . . , . . . [.] // -
 . – 2020. – . 27, 1. – . 85–95.
96. : -
 / . . . , . . . ,
 . . . [.] // . – 2015. –
 . 64, 4. – . 69–77.
97. ® (Oestrogel) // VIDAL: -
 . :
 www.vidal.ru/drugs/oestrogel__23864 (: 01.03.2018).
98. , . . . « »
 : / . . . ,
 . . . , . . . // . – 2019. –
 9. – . 32–39.
99. -
 / . . . , . . . , . . .
 [.] // . – 2015. – . 86, 1. – . 25–32.
100. -
 / . . . , . . . , . . . [.] //
 . – 2021. – . 23, 1. – . 92–96.

101. . . . [.] // . – 2015. – 1–2 (20). – . 20–24.
102. [.] // . – 2020. – . 18, 8–9. – . 7–12.
103. // , . – 2020. – . 19, 4. – . 50–56.
104. A meta– analysis on the prophylactic use of microlide antibiotics for the prevention of disease exacerbations in patients with Chronic Obstructive Pulmonary Disease / E.Donath,A. Chaudhry,L.F. Hernandez-Aya,L. Lit // *Resp. Med.* – 2013. – Vol. 107, 9. – P. 1385–1392.
105. A prospective randomized controlled study (RCT) of intra–uterine administration of Granulocyte Colony–Stimulating Factor (G–CSF) before embryo–transfer on resistant thin endometrium in IVF cycles /R. Singh, M. Singh, A. Jindal, P.C. Jindal // *Hum. Reprod.* – 2015. – Vol. 30. – P. 280.
106. A randomized clinical trial of endometrial perfusion with granulocyte colony–stimulating factor in in vitro fertilization cycles: Impact on endometrial thickness and clinical pregnancy rates / D.H. Barad, Y. Yu, V.A. Kushnir [et al.] // *Fertil.Steril.* – 2014. – Vol. 101, 3. – P. 710–715.
107. Altered uterine contractility in women with chronic endometritis / V. Pinto, M. Matteo, R. Tinelli [et al.] // *Fertil. Steril.*– 2015.– Vol. 103, 4.– P. 1049–1052.

108. Alves, R. A Review of Platelet Rich Plasma: History, Biology, Mechanism of Action, and Classification / R. Alves, R. Grimalt // *Skin Appendage Dis.* – 2018. – Vol. 4, 1. – P. 18–24.
109. An in vitro evaluation of the anti-inflammatory effects of platelet-rich plasma, ketorolac, and methylprednisolone / A.D. Mazzocca, B.R. McCarthy, J. Intravia [et al.] // *Arthroscopy.* – 2013. – Vol. 29, 4. – P. 675–683.
110. Anghelache-Lupa cu, I. Effect on endometrium quality in infertility uterine infectious etiology. Correlations between inflammatory factors and microbial agents (*Chlamydia trachomatis*, *Mycoplasma hominis*, *Ureaplasma urealyticum*) / I. Anghelache-Lupa cu. – Ia i, 2013. – 34 p.
111. Antimicrobial activity of pure platelet-rich plasma against microorganisms isolated from oral cavity / L. Drago, M. Bortolin, C. Vassena [et al.] // *BMC Microbiol.* – 2013. – Vol. 13. – P. 47.
112. Arsenic trioxide inhibits transforming growth factor-beta1-induced fibroblast to myofibroblast differentiation in vitro and bleomycin induced lung fibrosis in vivo / F. Luo, Y. Zhuang, M.D. Sides [et al.] // *Respir. Res.* – 2014. – Vol. 15. – P. 51.
113. Autologous Intrauterine Platelet-Rich Plasma Instillation for Suboptimal Endometrium in Frozen Embryo Transfer Cycles: A Pilot Study / S.R. Tandulwadkar, M.V. Naralkar, A.D. Surana [et al.] // *J. Hum. Reprod. Sci.* – 2017. – Vol. 10, 3. – P. 208–212.
114. Autologous platelet-rich plasma promotes endometrial growth and improves pregnancy outcome during in vitro fertilization / Y. Chang, J. Li, Y. Chen [et al.] // *Int. J. Clin. Exp. Med.* – 2015. – Vol. 8, 1. – P. 1286–1290.
115. Breuss, J.M. VEGF-initiated angiogenesis and the uPA/uPAR system / J.M. Breuss, P. Uhrin // *Cell Adhes. Migrat.* – 2012. – Vol. 6, 6. – P. 535–540.
116. Can autologous platelet rich plasma expand endometrial thickness and improve pregnancy rate during frozen-thawed embryo transfer cycle? A random-

- ized clinical trial / M. Eftekhar, N. Neghab, E. Naghshineh, P. Khani // *Taiwan J. Obstet. Gynecol.* – 2018. – Vol. 57, 6. – P. 810–813.
117. Check, J.H. Failure to improve a thin endometrium in the late proliferative phase with uterine infusion of granulocyte– colony stimulating factor / J.H. Check, R. Cohen, J.K. Choe // *Clin. Exp. Obstet. Gynecol.* – 2014. – Vol. 41, 4. – P. 473–475.
118. Christiansen, . . Intravenous immunoglobulin treatment for secondary recurrent miscarriage: a randomised, double–blind, placebo–controlled trial / . . Christiansen, . . Larsen, . Egerup // *BJOG.* – 2014. – Vol. 122, 4. – P. 500–508.
119. Chronic endometritis and altered embryo implantation: a unified pathophysiological theory from a literature systematic review / G. Buzzaccarini, A. Vitagliano, A. Andrisani [et al.] // *J. Assist. Reprod. Genet.* – 2020. – Vol. 37, 12. – P. 2897–2911.
120. Chronic endometritis and infertility / H.J. Park, Y.S. Kim, T.K. Yoon, W.S. Lee // *Clin. Exp. Reprod. Med.* – 2016. – Vol. 43, 4. – P. 185–192.
121. Chronic Endometritis: Old Problem, Novel Insights and Future Challenges/ E. Puente, L. Alonso, A.S. Laganà[et al.] // *Int. J. Fertil. Steril.* –2020. – Vol. 13, 4. – P. 250–256.
122. Comparison of the prevalence of chronic endometritis as determined by means of different diagnostic methods in women with and without reproductive failure / Y.Liu, X.Chen, J. Huang [et al.] // *Fertil. Steril.* – 2018. – Vol. 109. – P. 832–839.
123. Complex assessment of the effect of glycosaminoglycans on the system of haemostasis in patients with polycystic ovary syndrome / I.A. Lapina, L.A. Ozolinya, Y.E. Dobrokhotova [et al.] // *Gynecol. Obstet. Perinatol.* – 2019. – Vol. 18, 5. – P. 35–41.

124. Composition of platelet-rich plasma gel: A Western blot analysis / M.P. Galindo, O.G. Avila, M.L. Torrecillas [et al.] // *J. Oral Sci. Rehabil.* –2016. – Vol. 2, 2. – P. 42–48.
125. Effect of 1,25(OH)₂ vitamin D₃ on cytokine production by endometrial cells of women with repeated implantation failure / S. Rajaei, M. Mirahmadian, M. Jeddi-Tehrani [et al.] // *Gynecol. Endocrinol.* – 2012. – Vol. 28, 11. – P. 906-11.
126. Effect of growth hormone on the endometrial and endometrial blood flow in frozen thawed embryo transfer / H. Wu, L. Li, M. Li, H. Yuan // *J. Reprod. Med.* – 2013. – Vol. 22. – P. 914–917.
127. Effect of ovarian endometrioma on uterine and ovarian blood flow in infertile women / A. El-Mazny, A. Kamel, W. Ramadan [et al.] // *Int. J. Womens Health.* – 2016. – 8. – P. 677-682.
128. Effects of autologous platelet-rich plasma on implantation and pregnancy in repeated implantation failure: A pilot study / L. Nazari, S. Salehpour, S. Hoseini [et al.] // *Int. J. Reprod. Biomed. (Yazd).* – 2016. – Vol. 14, 10. – P. 625–628.
129. Effects of growth hormone on pregnancy rates of patients with thin endometrium / N. Cui, A.M. Li, Z.Y. Luo [et al.] // *J. Endocrinol. Invest.* – 2019. – Vol. 42, 1. – P. 27–35.
130. Effects of icariin on the expression of ER, VEGF, and KDR in the endometrial cells of thin endometrium / A.W. Le, L. Shan, Z.H. Wang [et al.] // *Genet. Mol. Res. J.* – 2015. – Vol. 14, 3. – P. 11250–11258.
131. Effects of platelet-rich plasma on mesenchymal stem cells isolated from rat uterus / P. Vishnyakova, D. Artemova, A. Elchaninov [et al.] // *Peer J.* – 2020. – 8. – P. e10415.
132. Efficacy evaluation of low-dose aspirin in IVF/ICSI patients evidence from 13 RCTs. A systematic review and meta-analysis / L. Wang, X. Huang,

X. Li [et al.] // *Medicine*. – 2017. – Vol. 96, 37. – DOI: 10.1097/MD.00000000000007720.

133. Efficacy of intrauterine infusion of granulocyte colony stimulating factor on patients with history of implantation failure: a randomized control trial / M. Eftekhar, S. Miraj, M.F. Mojtahedi, N. Neghab // *Int. J. Reprod. BioMed.* – 2016. – Vol. 14, 11. – P. 687–690.

134. Encapsulated VEGF121–PLA microparticles promote angiogenesis in human endometrium stromal cells / S. Abraham, G. Sanjay, N.A. Majiyd, A. Chinnaiyah // *J. Genet. Eng. Biotechnol.* – 2021. – Vol. 19, 1. – P. 23.

135. Endometrial microbiota in infertile women with and without chronic endometritis as diagnosed using a quantitative and reference range–based method / Y. Liu, E.Y. Ko, K.K. Wong [et al.] // *Fertil. Steril.* – 2019. – Vol. 112, 4. – P. 707–717.

136. Endometrial Vascularization Characterized by Optical Coherence Tomography and Immunohistochemistry in Women Undergoing In Vitro Fertilization–Embryo Transfer Treatment / T.S.M. Law, W.C. Cheung, F. Wu [et al.] // *Medicina (Kaunas)*. – 2019. – Vol. 55, 4. – P. 81.

137. Evaluating chronic endometritis in women with recurrent implantation failure and recurrent pregnancy loss by hysteroscopy and immunohistochemistry / M. Zargar, M. Ghafourian, R. Nikbakht [et al.] // *J. Minim. Invasive Gynecol.* – 2020. – Vol. 27, 1. – P. 116–121.

138. Evaluating the role of endometrial colour Doppler dynamic tissue perfusion measurements in in vitro fertilisation success / S. Akgün Kavurmacı, G. ahin, A. Akdoğan [et al.] // *J. Obstet. Gynaecol.* – 2021. – Oct. 28. – P. 1–7. – DOI: 10.1080/01443615.2021.1960292.

139. Evaluation of granulocyte colony–stimulating factor effects on treatment–resistant thin endometrium in women undergoing in vitro fertilization / M. Kunicki, K. Lukaszuk, I. Woclawek–Potocka [et al.] // *BioMed. Res. Int.* – 2014. – Vol. 2014. – P. 913235.

140. Follicular HCG endometrium priming for IVF patients experiencing resisting thin endometrium. A proof of concept study / E.G. Papanikolaou, D. Kyrou, G. Zervakakou [et al.] // *J. Assist. Reprod. Genet.* – 2013. – Vol. 30, 10. – P. 1341–1345.
141. Functional endometrial polyps in infertile asymptomatic patients: a possible evolution of vascular changes secondary to endometritis / F.M. Carvalho, F.N. Aguiar, R. Tomioka [et al.] // *Eur. J. Obstet. Gynecol. Reprod. Biol.* – 2013. – Vol. 170, 1. – P. 152–156.
142. Gleicher, N. A pilot cohort study of granulocyte colony-stimulating factor in the treatment of unresponsive thin endometrium resistant to standard therapies / N. Gleicher, A. Kim, T. Michaeli [et al.] // *Hum. Reprod.* – 2013. – Vol. 28, 1. – P. 172-7.
143. Glycosaminoglycans, proteoglycans and sulodexide and the endothelium: biological roles and pharmacological effects / V. Masola, G. Zaza, M. Onisto [et al.] // *Int. Angiol.* – 2014. – Vol. 33, 3. – P. 243–254.
144. Granulocyte colonystimulating factor in repeated IVF failure, a randomized trial / A. Aleyasin, Z. Abediasl, A. Nazari, M. Sheikh // *Reproduction.* – 2016. – Vol. 151, 6. – P. 637–642.
145. Growth hormone intrauterine perfusion combined with replacement cycle in the treatment of non–response thin endometrium: report of 5 cases / H. Yu, S.Gao, H. Tang [et al.] // *Int. J. Clin. Exp. Med.* – 2016. – 9. – P. 11982–11989.
146. Hannan, N.J. Pre-eclampsia: Challenges for Nanomedicine Development in Pregnancy / N.J. Hannan, N. de Alwis, N.K. Binder // *Trends Mol. Med.* – 2021. – Vol. 27, 8. – P. 824-825.
147. Histology of micro polyps in chronic endometritis / L. Resta, M. Palumbo, R. Rossi [et al.] // *Histopathology.* – 2012. – Vol. 60, 4. – P. 670–674.

148. Icariin promotes angiogenic differentiation and prevents oxidative stress–induced autophagy in endothelial progenitor cells / Y. Tang, A. Jacobi, C. Vater [et al.] // *Stem. Cells.* – 2015. – Vol. 33, 6. – P. 1863–1877.
149. Immunohistochemical Evaluation of Chronic Endometritis by CD138, CD3 and CD20 / T.K. Sarpal, R. Bhagat, R.S. Punia [et al.] // *Ann. Pathol. Lab. Med.* – 2019. – Vol. 6, 8. – DOI: 10.21276/APALM.2625.
150. Influence of growth hormone supplementation in patients with thin endometrium undergoing frozen embryo transfer / L. Li, X.–X. Sun, J.–Y. Yang [et al.] // *Reprod. Develop. Med.* – 2019. – 3. – P. 49–53.
151. Inside the Endometrial Cell Signaling Subway: Mind the Gap(s) / S. Makieva, E. Giacomini, J. Ottolina [et al.] // *Int. J. Mol. Sci.* – 2018. – Vol. 19, 9. – P. 2477.
152. Intrauterine infusion of autologous platelet–rich plasma in women undergoing assisted reproduction: A systematic review and meta–analysis / A. Maleki–Hajiagha, M. Razavi, S. Rouholamin [et al.] // *J. Reprod. Immunol.* – 2020. – Vol. 137. – P. 103078.
153. Kannar, V. Evaluation of endometrium for chronic endometritis by using syndecan–1 in abnormal uterine bleeding / V. Kannar, H.K. Lingaiah, V. Sunita // *J. Lab. Phys.* – 2012. – Vol. 4, 2. – P. 69–73.
154. Kitaya, K. Effect of early endometriosis on ovarian reserve and reproductive outcome / K. Kitaya // *Front. Biosci. (Schol Ed).* – 2015. – Vol. 7, 1. – P. 40–5.
155. Lafontaine, L. Immunoscore assay for the immune classification of solid tumors: Technical aspects, improvements and clinical perspectives / L. Lafontaine, F. Marliot, J. Galon // *Methods Enzymol.* – 2020. – 636. – P. 109–128.
156. Lazzaroni, E. A pilot cohort study of granulocyte colony–stimulating factor in the treatment of unresponsive thin endometrium resistant to standard therapies / E. Lazzaroni, D.H. Barad // *Hum. Reprod.* – 2013. – Vol. 28, 1. – P. 172–177.

157. Molecular Mechanism of Mesenchyme Homeobox 1 in Transforming Growth Factor 1-Induced P311 Gene Transcription in Fibrosis / Z. Wei, C. Han, H. Li [et al.] // *Front. Mol. Biosci.* – 2020. – Vol. 7. – P. 59.
158. Mouhayar, Y. Obstetrical complications of thin endometrium in assisted reproductive technologies: a systematic review / Y. Mouhayar, J.M. Franasiak, F.I. Sharara // *J. Assist. Reprod. Genet.* – 2019. – Vol. 36, 4. – P. 607–611.
159. Outcome Study of Five Cases Receiving In Vitro Fertilization After Treatment of Intrauterine Platelet-Rich Plasma (PRP) for Chronic Endometritis / F. Li, Y. Cui, D. Zhao [et al.] // *Panminerva Med.* – 2021. – DOI: 10.23736/S0031-0808.20.04247-0.
160. Physiological and pathological angiogenesis in endometrium at the time of embryo implantation / X. Chen, G.C. W. Man, Y. Liu [et al.] // *Am. J. Reprod. Immunol.* – 2017. – Vol. 78. – P. 1–7.
161. Physiopathologic mechanisms involved in mare endometrosis / M.R. Rebordão, A. Galvão, A. Szóstek [et al.] // *Reprod. Domest. Anim.* – 2014. – Vol. 49, 4. – P. 82–87.
162. Platelet-Rich Plasma in the Management of Chronic Endometritis Treatment in Women with Reproductive Health Disorders / V.O. Sklyarova, K.L. Shatylovich, A.L. Filipyuk [et al.] // *Eur. J. Med. Health Sci.* – 2020. – Vol. 2, 6. – DOI:10.24018/ejmed.2020.2.6.560
163. Prevalence and confounders of chronic endometritis in premenopausal women with abnormal bleeding or reproductive failure / D. Song, X. Feng, Q. Zhang [et al.] // *Reprod. Biomed. Online.* – 2018. – Vol. 36. – P. 78–83.
164. Prevalence of chronic endometritis in repeated unexplained implantation failure and the IVF success rate after antibiotic therapy / E. Cicinelli, M. Matteo, R. Tinelli [et al.] // *Hum. Reprod.* – 2015. – Vol. 30, 2. – P. 323–330.
165. Prognostic profiles and the effectiveness of assisted conception: secondary analyses of individual patient data / N.M. Van den Boogaard,

- A.J. Bendsdorp, K. Oude Rengerink [et al.] //Hum. Reprod. Update. – 2014. – Vol. 20, 1. – P. 141–151.
166. Reappraisal of ischemic heart disease: fundamental role of coronary microvascular dysfunction in the pathogenesis of angina pectoris / J.C. Kaski, F. Crea, B.J. Gersh, P.G. Camici // Circulation. – 2018. – Vol. 138. – P. 1463–1480.
167. Reduced Transforming Growth Factor- Activity in the Endometrium of Women with Heavy Menstrual Bleeding / J.A. Maybin, L. Boswell, V.J. Young [et al.] // J. Clin. Endocrinol. Metab. – 2017. – Vol. 102, 4. – P. 1299–1308.
168. Regenerative therapy by endometrial mesenchymal stem cells in thin endometrium with repeated implantation failure. A novel strategy / A.E. Tersoglio, S. Tersoglio, D.R. Salatino [et al.] // JBRA Assist. Reprod. – 2020. – Vol. 24, 2. – P. 118–127.
169. Regulatory T cells protect from autoimmune arthritis during pregnancy / A. Munoz-Suano, M. Kallikourdis, M. Sarris, A.G. Betz // J. Autoimmun. – 2012. – Vol. 38, 2-3. – P. J103-8.
170. Review: chronic endometritis and its effect on reproduction / F. Kimura, A. Takebayashi, M. Ishida [et al.]// J. Obstet. Gynaecol. Res. – 2019. – Vol. 45. – P. 951–960.
171. Role of Vascular Endothelial Growth Factor (VEGF) in Human Embryo Implantation: Clinical Implications / X. Guo, H. Yi, T.C. Li [et al.] // Biomolecules. – 2021. – Vol. 11, 2. – P. 253.
172. Sanjabi, S. Regulation of the Immune Response by TGF- : From Conception to Autoimmunity and Infection / S. Sanjabi, S.A. Oh, M.O. Li // Cold Spring Harb. Persp. Biol. – 2017. – Vol. 9, 6. – P. a022236.
173. Scarpellini, F. G-CSF treatment improves IVF outcome in women with recurrent implantation failure in IVF / F. Scarpellini, M. Sbracia // J. Reprod. Immunol. – 2012. – Vol. 94. – P. 103.

174. Successful Implantation and Live Birth Following Autologous Platelet–Rich Plasma Treatment for a Patient With Recurrent Implantation Failure and Chronic Endometritis / K. Sfakianoudis, M. Simopoulou, N. Nitsos [et al.] // *In Vivo*. – 2019. – Vol. 33, 2. – P. 515–521.
175. Sulodexide Prevents Peritoneal Fibrosis by Downregulating the Expression of TGF– 1 and Its Signaling Pathway Molecules / Z. Duan, J. Yao, N. Duan[et al.] // *Evid. Based Complement Alternat. Med.* – 2021. – Vol. 2021. – DOI: 10.1155/2021/2052787.
176. Synthesis of prenylated flavonols and their potents as estrogen receptor modulator / Z. Tao, J. Liu, Y. Jiang [et al.] // *Sci. Rep.* – 2017. – Vol. 7, 1. – P. 12445.
177. The diagnosis of chronic endometritis in infertile asymptomatic women: a comparative study of histology, microbial cultures, hysteroscopy, and molecular microbiology / I. Moreno, E. Cicinelli, I. Garcia–Grau [et al.] // *Am. J. Obstet. Gynecol.* – 2018. – Vol. 218, 6. – P. 602.
178. The effect of icariin for infertile women with thin endometrium: A protocol for systematic review / J. Du, H. Lu, X. Yu [et al.] // *Medicine*. – 2020. – Vol. 99. – P. 12.
179. The effects of growth hormone on clinical outcomes after frozen – thawed embryo transfer / W. Xue–Mei, J. Hong, Z. Wen–Xiang, L. Yang // *Int. J. Gynecol. Obstet.* – 2016. – Vol. 133. – P. 347–350.
180. The impact of chronic endometritis on endometrial fibrosis and reproductive prognosis in patients with moderate and severe intrauterine adhesions: a prospective cohort study / L. Liu, H. Yang, Y. Guo [et al.] // *Fertil. Steril.* – 2019. – Vol. 111, 5. – P. 1002–1010.
181. The measurement of endometrial volume and sub–endometrial vascularity to replace the traditional endometrial thickness as predictors of in–vitro fertilization success / A.M. Maged, A.M. Kamel, F. Abu–Hamila [et al.] // *Gynecol. Endocrinol.* – 2019. – Vol. 35, 11. – P. 949–954.

182. The reliability of the histological diagnosis of endometritis in asymptomatic IVF cases: a multicenter observer study / J.C. Kasius, F.J.M. Broekmans, D.M.D.S. Sie-Go [et al.] // *Hum. Reprod.* – 2012. – Vol. 27. – P. 153–158.
183. The role of G-CSF in recurrent implantation failure: a randomized double blind placebo control trial / F. Davari-Tanha, E.S. Tehraninejad, M. Ghazi, Z. Shahraki // *Int. J. Reprod. Bio-Med.* – 2016. – Vol. 14, 12. – P. 737–742.
184. Treatment of thin endometrium with autologous platelet-rich plasma: a pilot study / S. Zadehmodarres, S. Salehpour, N. Saharkhiz, L. Nazari // *JBRA Assist. Reprod.* – 2017. – Vol. 21, 1. – P. 54–56.
185. Unified diagnostic criteria for chronic endometritis at fluid hysteroscopy: proposal and reliability evaluation through an international randomized-controlled observer study / E. Cicinelli, A. Vitagliano, A. Kumar [et al.] // *Fertil. Steril.* – 2019. – Vol. 112, 1. – P. 162–173.
186. Use of Platelet-Rich Plasma Plus Suramin, an Antifibrotic Agent, to Improve Muscle Healing After Injuries / W.C. Tsai, T.Y. Yu, G.J. Chang [et al.] // *Am. J. Sports Med.* – 2021. – Vol. 49, 11. – P. 3102–3112.
187. Using autologous intrauterine plateletrich plasma to improve the reproductive outcomes of women with recurrent implantation failure / F. Aghajanzadeh, S. Esmailzadeh, Z. Basirat [et al.] // *JBRA Assist. Reprod.* – 2020. – Vol. 24, 1. – P. 30–33.
188. VEGF may contribute to macrophage recruitment and M2 polarization in the decidua / K.C. Wheeler, M.K. Jena, B.S. Pradhan [et al.] // *PLoS One.* – 2018. – Vol. 13, 1. – DOI: 10.1371/journal.pone.0191040.